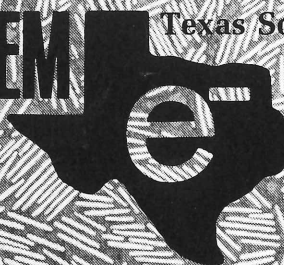


TSEM



Texas Society for Electron Microscopy

JOURNAL
VOLUME 24, NUMBER 1, 1993
ISSN 0196-5662

INTRODUCING

ISIS

LINK ISIS - Microanalysis made easy

ISIS

ISIS



ISIS

ISIS

**All we know
about microanalysis
- at your fingertips.**

The Link Isis X-ray microanalysis system is dedicated to making routine microanalysis easy.

Combining 20 years of experience in high precision microanalysis with the intuitive simplicity of the Microsoft Windows user interface, the Link Isis system allows even the occasional user to achieve high quality results, quickly. Productivity is boosted for both dedicated and occasional users with fast data acquisition, reduced learning times and rapid report generation.

- ▲ Innovative system architecture for the fastest microanalysis available.
- ▲ BEAMSEARCH software for point and see analysis.
- ▲ Simple, accurate peak identification software.
- ▲ Full microscope automation for high productivity.
- ▲ LABBOOK database for easy, efficient data management.
- ▲ LINK PENTAFET PLUS detectors for the highest levels of accuracy.

Add to these our Total Quality manufacturing environment, comprehensive training programs, telephone hotline service and technical support facilities around the world. The result is the finest value-for-money X-ray microanalysis system available today.

OXFORD

Oxford Instruments, Microanalysis group

601 Oak Ridge Turnpike
Oak Ridge, TN 37830
USA
Tel 615 483 8405 Fax 615 483 5891

ISIS

Offices in USA - Foster City 415 5780202; Boston 508 369 8850; Canada 416 568 9188; Australia 2-875 3130; France 1 69 41 8990; Germany 8121 81094; Sweden 8 767 9170; agents throughout the world.



TSEM OFFICERS 1993-1994

President:

HAL K. HAWKINS
Department of Pathology
U.T. Medical Branch
11th & Texas
Galveston, Texas 77555-0609
(409) 770-6655 FAX (409) 772-2500

President Elect:

NANCY K. R. SMITH
Cellular and Structural Biology
Univ. of Texas Health Science Center
7703 Floyd Curl Drive
San Antonio, Texas 78284-7762
(512) 567-3861 FAX (512) 567-2490

Past President:

LYNN D. GRAY
Department of Cell Bio. & Enviro. Science
Univ. of Texas Health Science Center
P.O. Box 2003
Tyler, Texas 75710
(903) 877-7575 FAX (903) 877-7558

Secretary:

KEITH R. FRY
Center for Biotechnology
Baylor College of Medicine
4000 Research Forest Drive
The Woodlands, Texas 77381
(713) 363-8415 FAX (713) 363-8475

Secretary Elect:

SUSAN E. ROBBINS
Department of Pathology, Room 212-B
Baylor College of Medicine
1200 Moursund
Houston, Texas 77030
(713) 798-4658 FAX (713) 798-5838

Treasurer:

CAROLYN CORN
Department of Cell Bio. & Enviro. Science
Univ. of Texas Health Science Center
P.O. Box 2003
Tyler, Texas 75710
(903) 877-7575 FAX (903) 877-7558

Program Chairman:

PAULA WILLIAMSON
Department of Biology
Southwest Texas State University
San Marcos, Texas 78666
(512) 245-2171 FAX (512) 245-8095

Program Chairman Elect:

MITCHELL D. McCARTNEY
EM Unit, RO-11
Alcon Laboratories, Inc.
6201 South Freeway
Ft. Worth, Texas 76134-2099
(817) 551-4620 FAX (817) 551-4584

APPOINTED OFFICERS

Corporate Member Representative:

TONY L. CARPENTER
Carl Zeiss, Inc.
6809 Ragan Drive
The Colony, Texas 75056
(214) 625-8525 FAX (914) 681-7443

Student Representative:

DAVID C. GARRETT
Department of Biology
Univ. of Texas at Arlington
P.O. Box 19498
Arlington, Texas 76019
(817) 273-2871 FAX (817) 273-2855

TSEM Journal Editor:

LOUIS H. BRAGG
Department of Biology
Univ. of Texas at Arlington
P.O. Box 19498
Arlington, Texas 76019
(817) 273-2402 FAX (817) 273-2855

Contents

TEXAS SOCIETY FOR ELECTRON MICROSCOPY JOURNAL VOLUME 24, NUMBER 1, 1993 ISSN 0196-5662

Louis H. Bragg, Editor

Department of Biology, The University of Texas at Arlington, Arlington, TX 76019

Texas Society for Electron Microscopy

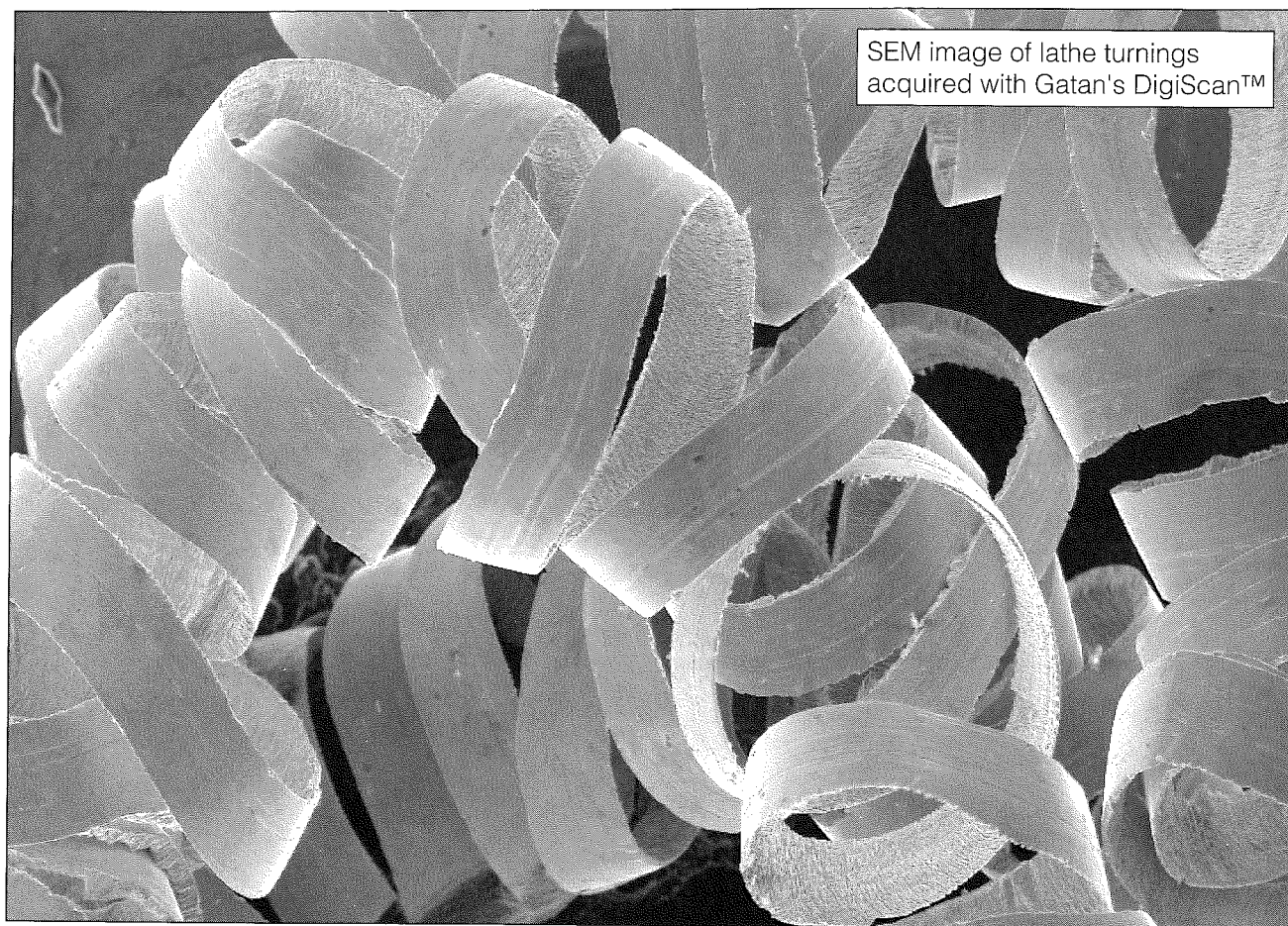
"For the purpose of dissemination of research with the electron microscope."

President's Message	5
Treasurer's Report	7
Join EMSA	11
EMSA Application for Membership	15
Certification Board Examinations for Electron Microscopy Technologist	17
Calendar of Meetings	19
EMSA Electronic Bulletin Board System	21
Advertisers' Index	23
Editorial Policy	23
TSEM Student Competition	25
TSEM Student Competition Application	26
Information for Authors	27
MSA Undergraduate Scholarship Program	29
MSA Undergraduate Scholarship Application	31
Abstracts	33
Answer to "What Is It?"	36
TSEM Application For Membership	37
TSEM Members	41
Corporate Members	45
What Is It?	Back Cover

ON THE COVER

Transmission electron micrograph of the tapetum lucidum of the Bay Anchovy. Magnification = 4950X. Photo — Howard J. Arnott, Biology Department, The University of Texas at Arlington, 76019.

And you thought we only made great products for your TEM



SEM image of lathe turnings
acquired with Gatan's DigiScan™

Introducing GATAN's Point-and-Click SEM Image Acquisition System

GATAN's DigiScan™ is a flexible system for acquiring large dynamic range SEM or STEM images with Macintosh computers. It opens up all the possibilities of desktop digital imaging for your existing scanning electron microscopes.

Major features include:

- image sizes up to 4k x 4k pixels
- on-line image rotation
- real-time imaging (up to 2 million pixels/sec.)
- variable time integration at each pixel
- simultaneous viewing of up to 4 analog or digital signals
- analog signal conversion with up to 16-bit precision
- pulse counting at >10 MHz
- powerful image processing provided by GATAN's DigitalMicrograph™ software
- image archiving
- cut-and-paste transfer of images into reports
- draft copy printing on laser printers and high-quality printing on continuous tone printers
- image transfer via Ethernet

Gatan Inc.

6678 Owens Drive, Pleasanton, CA 94588-3334, USA
780 Commonwealth Drive, Warrendale, PA 15086-7598, USA
Ingolstädter Straße 40, D-8000 München 45, Germany
3 Saxon Way E, Oakley Hay, Corby, Northants, NN18 9EY, UK

**Gatan GmbH
Gatan Ltd.**

Tel 510 463 0200
Tel 412 776 5260
Tel 089 35 23 74
Tel 0536 743150



President's Message

I can hardly believe the year has gone by so fast! As my term as President draws to a close, I want to thank the members of TSEM for allowing me the privilege of representing you. I also want to thank the officers for all of their help and hard work in putting the meetings together, getting mailouts to the membership, keeping the finances straight and producing a quality *Journal* — plus taking care of all the many details that are necessary evils of running a scientific society. Dealing with hotels and planning a quality scientific program for each meeting are tremendous jobs which involve **all** of the elected and appointed officers to some extent. I hope we've done a good job for you. Along that line, I want to encourage the membership to take an **ACTIVE** role in **your** Society. Let the officers know what you want in the way of programs, special events, symposia, workshops, etc. This benefits everyone and keeps us in line with the latest scientific developments. By the way, there is a lot of untapped talent out there in the membership — run for an office and make a difference!

Along a more serious line, we in TSEM owe a huge debt of gratitude to Dr. Wayne Sampson for his efforts on behalf of the Society regarding our state and federal tax exempt status. Wayne has spent a great deal of his personal time and effort in recruiting and working with attorney Michael Middleton to get the voluminous required paperwork re-filed with

appropriate state agencies and the IRS. We also appreciate Mr. Middleton handling our affairs in such a timely and professional manner. Many thanks to both of these individuals for a job above and beyond.

Please note that this issue of the *Journal* contains the latest membership list. If you would, take a minute and look it over. If changes need to be made, please let Keith Fry (Secretary) know so that he can update the data base. We don't want to miss anyone!

I look forward to seeing you in Corpus Christi. We have an excellent program planned and Corpus is always a great place to meet. The Fall 1993 meeting will be at the Tremont House in Galveston, so plan ahead and keep thinking, "No hurricanes . . . no hurricanes . . ."!

Finally, I wish Hal Hawkins the best of luck in the coming year as he serves as TSEM President. I know everyone will give him their support. Remember, we need your abstracts for the meetings and your papers for the *Journal*. My best to all, I've enjoyed the year!

Sincerely,

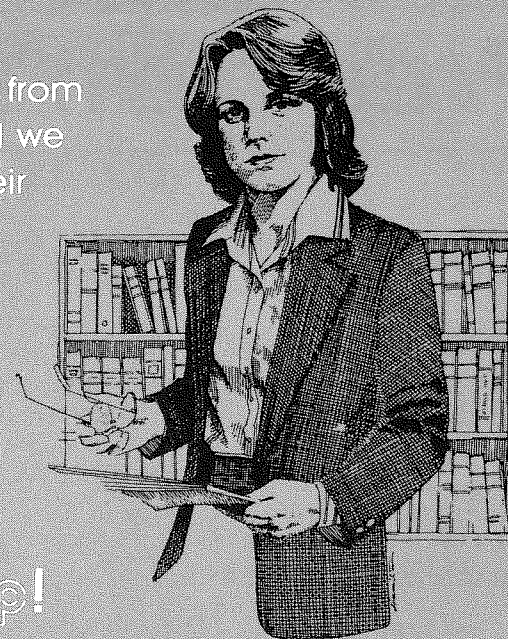
Lynn D. Gray, Ph.D.
President, 1992-1993

V.A. & Federal
Customers: Order
on GSA #00F-4746A

"We bought our NEW Diamond Knives from
AMCorp and a year later we're glad we
did. Their quality is world class and their
warranty is unbeatable."

ISN'T IT TIME FOR YOU TO
CONSIDER INVESTING IN
ANOTHER PROVEN QUALITY
SOURCE FOR YOUR NEW
DIAMOND KNIVES? . . . **AMCorp!**

for Reichert, Sorvall, LKB, DuPont and RMC



WARRANTY

- **1 Year Unconditional Warranty**
- **Lifetime Resharpener Guaranteed**
- **Lifetime Free Diamond Replacement**
- **Satisfaction Guaranteed**
(request copy of warranty and specifications)

Of Course We:

— Use Quality Gemstones — Provide Long Lasting Durable Edges: 1-10mm — Accept Trade-Ins — Ship from Stock — Supply Any Style Boat Free —

— Resharpen

- | | | | |
|-------------|----------|------------|--------------|
| • Diatome | • DuPont | • Jumdi | • Sugg |
| • DDK | • IVIC | • Ge Fe Ri | • Rondikn |
| • MicroStar | • W.R. | • Sag | • LKB |
| • Diatech | • MJO | • Dehmer | • and others |

Stainless Steel, Cryo-Dry, Cryo-Wet, DuPont or Superboat™
Holders On Request, No Charge

FREE: 1-800-528-5252 MA: 1-617-965-6340 FAX: 1-617-964-1052



• PO Box 67285 • Chestnut Hill, MA 02167 • 1-800-528-5252 / 1-617-965-6340 (MA)

Quality is remembered long after price is forgotten.

Treasurer's Report

TREASURER'S REPORT For Period Ending December 31, 1992

ASSETS ON JANUARY 1, 1992:

Certificate of Deposit No. 113515 (formerly CD#177576)	\$3,455.96
Certificate of Deposit No. 2414483036 (formerly CD#0014483036)	1,612.50
Checking Account No. 44059412	5,872.62
TOTAL	\$10,941.08

CHECKING ACCOUNT RECEIPTS:

Dues	\$4,783.00
Spring 1992 Meeting Registration	1,520.00
Workshop	430.00
Exhibitors	925.00
Donations and Grants	850.00
Guest	320.00
Fall 1992 Meeting Registration	1,080.00
Exhibitors	1,705.00
Workshop	290.00
Donations and Grants	180.00
Guest	180.00
Journal Advertisements 23:1	1,995.00
23:2	1,750
TOTAL	\$16,273.00
Certificate of Deposit Interest	381.05

EXPENSES:

Journal, Postage	\$3,548.56
Stationary and Office Supplies	84.05
Mailouts	1,583.57
Spring 1992 Meeting	4,504.53
Workshop	513.00
Student Competition/Travel	135.00
Fall 1992 Meeting	3,635.28
Workshop	206.22
Student Travel	295.00
Miscellaneous	254.10
Tax and Legal Fees	1,617.65
Checking Account Service Charge	84.40
TOTAL	\$16,744.05
Certificate of Deposit No. 9005997	4,000.00

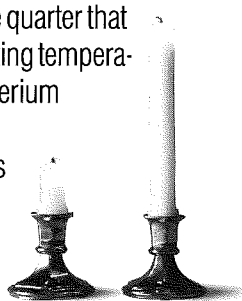
ASSETS AS OF DECEMBER 31, 1992:

Certificate of Deposit No. 113515 (formerly CD#177576)	\$3,651.24
Certificate of Deposit No. 2414483036 (formerly CD#0014483036)	1,688.30
Certificate of Deposit No. 9005997	4,109.97
Checking Account No. 44059412	1,401.57
TOTAL	\$10,851.08

Five Good Reasons To Switch To Our New CeBix™ Cathode (And One Silly One)

1. Longer Life.

With an evaporation rate one quarter that of LaB₆ cathodes at average working temperatures (1750 K), FEI's new CeBix Cerium Hexaboride Mini-Vogel Mount cathodes can last up to four times longer. And require less frequent wehnelt cleaning.



2. High Brightness.

Like the thousands of FEI LaB₆ cathodes now in daily use, our new CeBix cathodes are



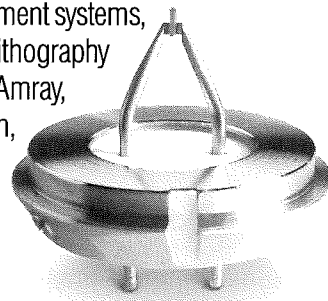
an order of magnitude brighter than tungsten cathodes. The advantage of CeBix cathodes is they give you that brightness much longer.

3. More Resistant To Carbon Contamination.

Studies show CeBix cathodes desorb carbon contamination faster, allowing the emission current to stabilize much more quickly than with LaB₆ cathodes.

4. Perfect Fit.

Mounts are available for most popular SEMs and TEMs, CD measurement systems, and Auger and E-Beam lithography systems. These include Amray, Camscan, Hitachi, Noran, Topcon/ISI, JEOL, Leica/CI, Perkin-Elmer PHI, Philips and Zeiss.



5. Better Value.

With their lower evaporation rate and consequent longer life, CeBix cathodes have a much lower cost per operating hour than even our own long-lasting LaB₆ cathodes.

6. Free Mug.

If you need another reason to try our new CeBix cathode, here it is. The first 500 purchasers will receive a special, heat-sensitive mug that transforms as you fill it. See, we told you there would be a silly reason.



Call to order or for more information.



FEI Company
19500 N.W. Gibbs Drive, Suite 100
Beaverton, OR 97006-6907
(503) 690-1500, FAX: (503) 690-1509



Partners in
Discovery:

Electron Microscopes
from Carl Zeiss

The New Era of OMEGA Filter Electron Microscopy Has Begun

Why use yesterday's technology to face today's challenges? Discover the Zeiss OMEGA Filter and lift the veil which blurs your TEM vision. The revolutionary OMEGA Filter offers you a clear choice.

Filtered Imaging

Images of unprecedented brilliance are the hallmark of the new EM 912 OMEGA for any kind of specimen.

Three dimensional reconstructions and stereo imaging of thick specimens become routine.

Filtered Diffraction

Filtered diffraction patterns are the clear future for diffraction analysis. With the Zeiss EM 912 OMEGA's fully motorized, five axis goniometer and specimen position recall, energy filtered CBED and selected area diffrac-

tion zone axes are a push button away. Diffraction pattern analyses and measurements have never been so easy.

Elemental Imaging

Zeiss continues to be the clear leader in this field. Element specific images from the full magnification range are at your fingertips.

Elemental X-ray Analysis

Designed to provide the highest X-ray detection efficiency, a 1.6 nm probe diameter, and the lowest background (hole count), the new EM 912 OMEGA with the integrated OMEGA Filter is designed to be the new standard for analytical performance. The advantages of OMEGA Filter integration are obvious when you sit down at the instrument! ...And when you see the results!

Carl Zeiss, Inc.
Electron Optics Division
Thornwood, NY 10594
(914) 681-7745
Fax (914) 681-7443



Carl Zeiss
Electron Optics Division
D-7082 Oberkochen, Germany
(7364) 20 2700
Fax (7364) 20 4530

Electron Microscopy Sciences

Building a Solid Foundation
of Commitment in the
Microscopy Community.

The chemicals and supplies
you want; the quality, value,
and service you need.

Many new products, as well as never
before seen items have been added
to our already extensive line of the
highest quality chemicals, supplies,
accessories, and equipment filling
all of your microscopy needs.

Call or write today for a copy of our newest catalog.

321 Morris Road • Box 251 • Fort Washington, PA 19034

Toll-free: 1-800-523-5874 • (215) 646-1566 • Fax: (215) 646-8931 • Telex: 510-661-3280

Join EMSA

Electron Microscopy Society of America

— ENJOY THESE ADVANTAGES —

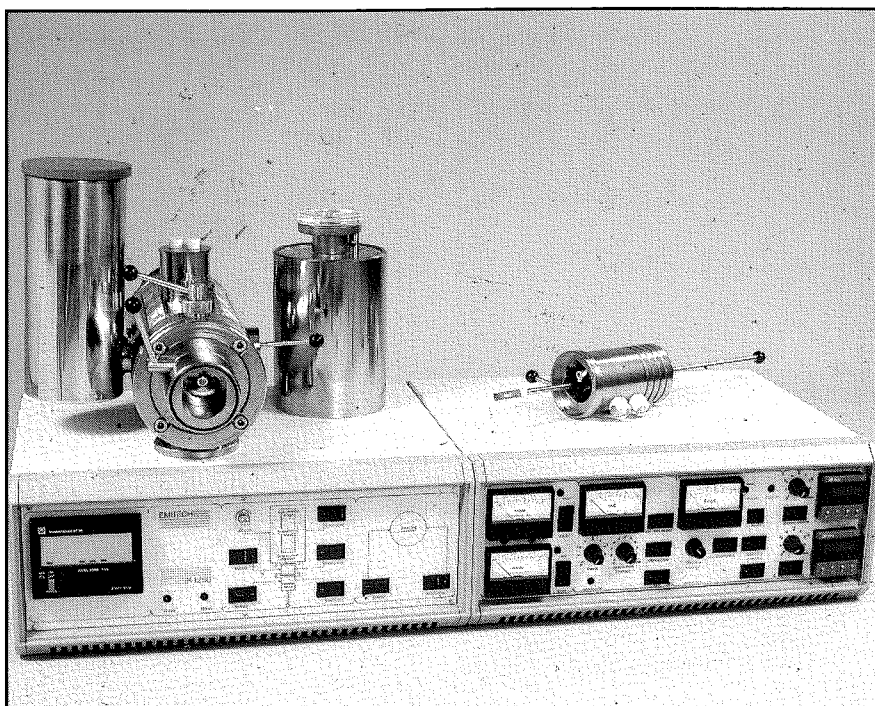
- ▶ EMSA Bulletin twice each year
 - ▶ Special subscription rates on microscopy journals
 - ▶ Employer/employee listings
 - ▶ AV Tape Rental and Copying Privileges
 - ▶ Microscopy Book List
 - ▶ Poster Exhibit Use
 - ▶ Tutorials and Demos at National Meeting
 - ▶ Listing of Short Courses in Microscopy
 - ▶ Instrument Instructions Clearing House
 - ▶ Listing of Microscopy Software and other AVs
 - ▶ Annual meeting that covers all areas of microscopy with exhibits of current instruments & supplies
- and**
- ▶ Lots of new friends as microscopists are great people and friendly!

Write for a membership form

EMSA

P.O. Box EMSA — Woods Hole, MA 02543
(617) 540-7639

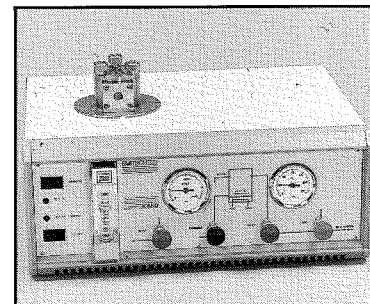
EMITECH ≡ K SERIES



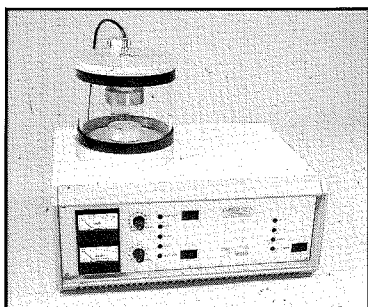
The K1250 'TURBO' Cryogenic Preparation System



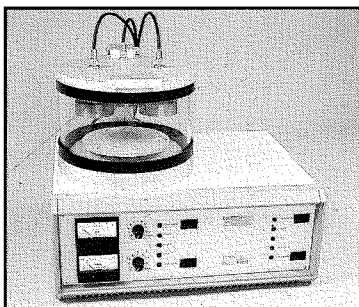
The K750 Freeze Drier



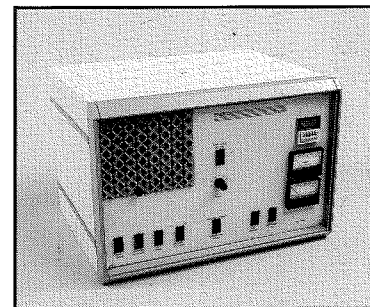
The K850 Critical Point Drier



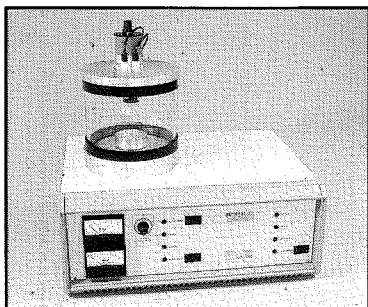
The K550 Automatic Coater



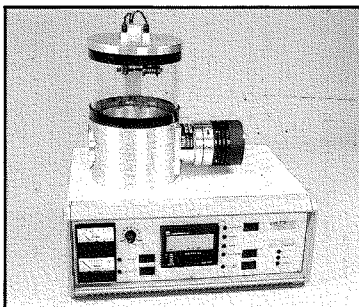
The K650 Large Sample Coater



The K1050 Plasma Asher



The K450 Carbon Coater



The K950 Turbo Evaporator

OTHER PRODUCTS

(not illustrated)

The K250 Carbon Coating

Attachment

The K350 Sputter Coating

Attachment

A Consumables Catalogue with a range of items in everyday use in the Laboratory

U.S.A.

EMITECH INC

Tel: 713-893-2067 Fax: 713-893-8443

UNITED KINGDOM

EMITECH LTD

Tel: 0233 646332 Fax: 0233 640744

BENELUX

J. J. BOS, BV.

Tel: 01828-19333 Fax: 01828-11770

CANADA

CANBERRA-PACKARD CANADA LTD

Tel: (416) 890 6422

Fax: (416) 890 6466

FRANCE

JOSE DELVILLE

Tel: (1) 34516211 Fax: (1) 30 61 49 57

GERMANY

RÖNTGENANALYTIK MESSTECHNIK

Tel: 06128 71080 Fax: 06128 73601

ITALY

2M STRUMENTI Srl

Tel: 06/8895319 Fax: 06/8895241

KOREA

RIGONG INTERNATIONAL INC

Tel: 82-2-719-7850 Fax: 82-2-718-5362

SPAIN

MICROSCOPIA Y ANALITICA, SA.

Tel: (91) 890 7606 Fax: (91) 890 7610

SWEDEN

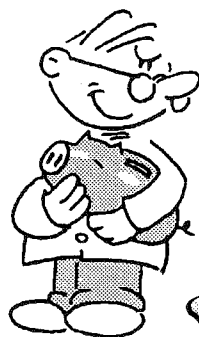
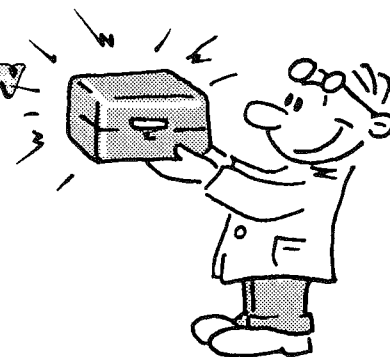
LEICA AB.

Tel: (08) 7513340 Fax: (08) 7506010

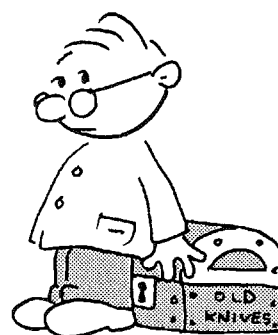
SERVING THE SCIENCE OF ELECTRON MICROSCOPY

LOOK, A deal you may not want to pass:
Exchange any old diamond knife for a
new MICRO STAR at the resharpener price.

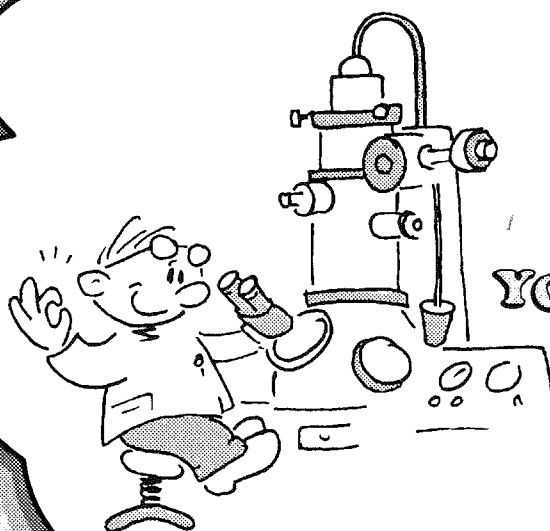
**YOU GET A BRAND NEW
MICRO STAR
DIAMOND KNIFE**



**YOU KEEP
YOUR MONEY**



**YOU KEEP
YOUR
EXCHANGE KNIFE**



**UNTIL YOU VERIFY
YOUR NEW MICRO STAR'S
TOP QUALITY**

To order give us the size and serial number of the knife you are exchanging. We accept any brand, any type, any size. In a few days you will receive a new MICRO STAR diamond knife the same size as your exchange.

Keep your knife and do not authorize payment until you have tested the new MICRO STAR and are totally satisfied with its top quality. Please test it within a month. Sample prices: 2mm \$890, 3mm \$1090. Call us for full information.

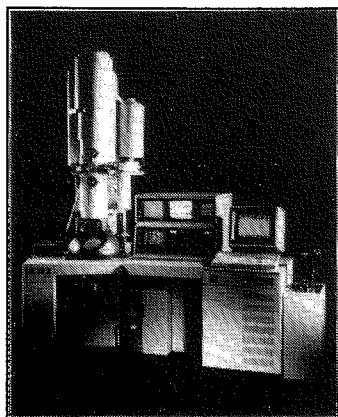
 **micro engineering inc.**

RT 2 BOX 474, HUNTSVILLE TX 77340
TEL 409 291 6891 FAX 409 294 9861

800 533 2509

SCANNING & TRANSMISSION ELECTRON MICROSCOPES

The XL Series of Scanning Electron Microscopes combines superb electron optics (2.0nm, LaB6, XL30), optimum analytical geometry (a common 10mm eucentric point and working distance for EDS) with Microsoft® Windows™ control for unprecedented ease-of-use. Including digital imaging with standard TIFF file format, image archiving and LAN compatibility, the XL Series offers performance and productivity at a competitive price.



The CM Series of Transmission Electron Microscopes, including the CM20-UltraTWIN (point resolution 0.19 nm at 200kV with $\pm 20^\circ$ tilt), CM20 FEG with its Schottky field emitter and CM12CRYO for beam sensitive specimens provides the optimum system for your materials and life science applications. Featuring the unique TWIN lens and eucentric goniometer, these microscopes combine imaging, diffraction and analytical modes at the highest spatial resolution and detection efficiency, together with unequalled ease-of-use.

You get superior performance. And, with these software based systems you get the latest technology. Right for today. Ready for tomorrow. Get the Philips Electron Optics team working for you.

Philips Electronic Instruments, 85 McKee Drive, Mahwah, NJ 07430; 201-529-3800; (fax) 201-529-2252



PHILIPS

ELECTRON MICROSCOPY SOCIETY OF AMERICA

Box EMSA, Woods Hole, MA 02543 • (508) 540-7639 or (800) 538-EMSA

APPLICATION FOR MEMBERSHIP

Name (*print*): _____ Dr. ☐ Mr. ☐ Ms. ☐

Institutional Affiliation: _____

Mailing Address: _____

Phone (*days*): () _____ Major Interest: Physical Sciences ☐ Biological Sciences ☐

Fax: () _____ E-Mail: () _____

Signature of nominating EMSA Member: _____

Signature of advisor (*for student applicants*): _____

Signature of applicant: _____ Date: _____

EMSA Local Affiliate Societies (*Choose one*)

- | | |
|---|--|
| <input type="checkbox"/> Alabama Electron Microscopy Society | <input type="checkbox"/> New England Society for Electron Microscopy |
| <input type="checkbox"/> Appalachian Regional Electron Microscope Society | <input type="checkbox"/> New York Society for Electron Microscopy |
| <input type="checkbox"/> Arizona Society for Electron Microscopy and Microbeam Analysis | <input type="checkbox"/> Northern California Society for Electron Microscopy |
| <input type="checkbox"/> Central States Electron Microscopy Society | <input type="checkbox"/> North Carolina Society for Electron Microscopy and Microbeam Analysis |
| <input type="checkbox"/> Chesapeake Society for Electron Microscopy | <input type="checkbox"/> Northwestern Ohio Electron Microscopy Society |
| <input type="checkbox"/> Connecticut Electron Microscopy Society | <input type="checkbox"/> Oklahoma Society for Electron Microscopy |
| <input type="checkbox"/> Electron Microscopy Society of Northeastern Ohio | <input type="checkbox"/> Pacific Northwest Electron Microscopy Society |
| <input type="checkbox"/> Electron Microscopy Society of the Ohio River Valley | <input type="checkbox"/> Philadelphia Electron Microscopy Society |
| <input type="checkbox"/> Florida Society for Electron Microscopy | <input type="checkbox"/> San Diego Society for Electron Microscopy |
| <input type="checkbox"/> Iowa Microbeam Society | <input type="checkbox"/> Southern California Society for Electron Microscopy |
| <input type="checkbox"/> Louisiana Society for Electron Microscopy | <input type="checkbox"/> Southern California Society for Electron Microscopy Technologists |
| <input type="checkbox"/> Michigan Electron Microscopy Forum | <input type="checkbox"/> South Carolina Society for Electron Microscopy |
| <input type="checkbox"/> Midwest Society of Electron Microscopists | <input type="checkbox"/> Southeastern Electron Microscopy Society |
| <input type="checkbox"/> Minnesota Electron Microscopy Society | <input type="checkbox"/> Texas Society for Electron Microscopy |
| <input type="checkbox"/> Mountain States Society for Electron Microscopy | |

Enclose a check (*U.S. funds, drawn on a U.S. bank, or International Money Order*) for one year's dues, payable to EMSA, and a brief statement of your qualifications, experience, and/or student status.

Regular Member: \$30 ☐

Student Member: \$5 ☐

DIATOME U.S.

The Diatome Collection...

The Leading Edge in Diamond Knife Technology

Over the years Diatome has been the innovator in Diamond Knife Technology. We were the first company to perfect and introduce low angle diamond knives, as well as diamond knives for cryo ultramicrotomy and light microscopy.

We are constantly researching and developing new products that we believe will give our customers the results they require from their work. Now included in that list is our Diamond Trimming Tool and "Static-Line" Ionizer.

Whether your needs are Biological or Materials related, in EM or LM, at ambient or low temperatures, Diatome has the answer. With three different angles (35°, 45°, 55°) and 6 different types of knives (ultrathin, semithin, cryo-wet, cryo-dry, histo, histo-cryo), covering the entire microscopy spectrum.

Diatome is committed to your satisfaction. If you have any special requirements, or need assistance in choosing a knife for your special application just contact us with your needs. Our technical staff, upon receipt of your sample block, will do all necessary testing and make their recommendations to you. Custom boats (color and shape) and custom knives for special applications are available as well.

All new knives are shipped within 4 weeks if they are not immediately available from stock (we do keep an extensive stock of knives on hand for immediate delivery).

For a perfect cut every time look for the name that is leading the way - DIATOME. Your guarantee for accuracy, durability, and the highest standard in quality.

DIATOME U.S.

Call or write for our complete set of literature today.

DIATOME U.S.:

P.O. Box 125, Fort Washington, PA 19034

Telephone: 215-646-1478 • Fax: 215-646-8931

ELECTRON MICROSCOPY SOCIETY OF AMERICA CERTIFICATION BOARD EXAMINATIONS

ELECTRON MICROSCOPY TECHNOLOGIST

—(BIOLOGICAL SCIENCES)—

GENERAL ELIGIBILITY REQUIREMENTS:

1. Membership in EMSA.
2. ONE of the following conditions must be met:
 - 2 years (60 credits) college or equivalent, including science and TEM (1 year laboratory) courses; science courses to include one each of chemistry, physics and biology; math through trigonometry
 - 1 year (30 credits) college or equivalent, including one course each of chemistry and physics, and 1 year of recent full-time work experience (within the past 5 years) in a TEM laboratory
 - high school diploma and 2 years of recent full-time work experience in a TEM laboratory
 - 3 years of recent full-time work experience in a TEM laboratory
 - 6 years full-time TEM work experience within the past 8 years.

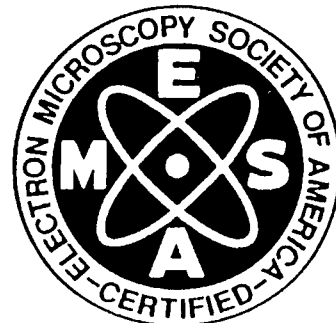
IMPORTANT DEADLINES:

Examinations are administered twice a year (two cycles per year).

Deadlines for receipt of applications are: October 1 and April 4.

FOR APPLICATIONS AND ADDITIONAL INFORMATION:

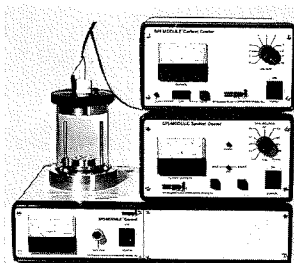
MSA CERTIFICATION OFFICE
MSA BUSINESS OFFICE
P.O. BOX MSA
WOODS HOLE, MA 02543



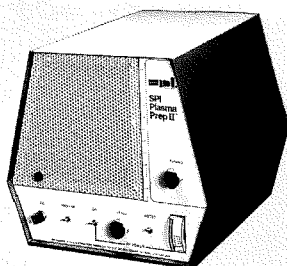
SPI [®] **is the Source**

SUPPLIES for the **BEST** in Electron Microscopy Supplies

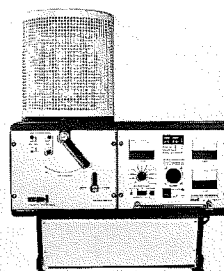
INSTRUMENTS



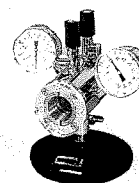
SPI-Module™
Sputter/Carbon
Coating Systems



Plasma Prep II™
Low Temperature
Etcher/Asher



Vacu-Prep™
Benchtop
Vacuum Evaporator



SPI-CPD™
Critical Point
Dryer

CHEMICALS

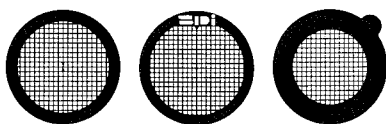
**Buffers
Fixatives & Stains
Embedding Media**
including:

- Osmium Tetroxide
- LR White®/LR Gold®
- Glutaraldehyde
- Cacodylic Acid

SPI-MARK™ COLLOIDAL GOLD REAGENTS

- Highest Purity
- Select from:
 - Protein A-gold
 - Protein G-gold
 - EM and LM Conjugates

GRIDS



- Over 200 styles
- Quantity discounts
- Custom Coating
- Regular
- Slim Bar™
- SuperGrids™

TWEEZERS



Over 200 Styles!

- Stainless Steel
- Teflon® coated
- Gold Plated
- 100% Antimagnetic
- Miracle Tip™ w/repairable tips
- SPI-Swiss™ & Dumont™ brands

BOOKS & LEARNING AIDS



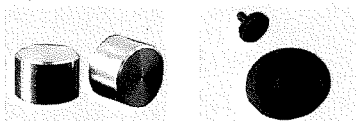
Over 100 titles
in our
Microscopists'
Library!

LAB CLEANING SUPPLIES



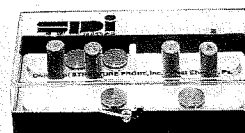
- O.K. Duster & O.K. Easy Duster
Contain no CFC's
- Bell Bright™/Metal Bright™
- SPI-Clean™ Cloths and Swabs

SPECIMEN MOUNTS



ALUMINUM - Lathe,
SPI-LUSTRE™, High Polish
CARBON - Standard,
Ultra Smooth™ Surface

STORAGE BOXES



- 5 types to match your mounts
- Desiccator-dry
- Clean, dust-free

ADHESIVES



- Conductive Silver Paints/Pastes
- Conductive Carbon Paints/Pastes
- SPI Supplies' own Silver Streaker™

Call Toll-Free: 1-800-2424-SPI • FAX: 1-215-436-5755



SPI Supplies Division of **STRUCTURE PROBE, INC.**
P.O.Box 656, West Chester, PA 19381-0656 USA



CALENDAR OF MEETINGS

SCANNING 93

April 21-23, 1993

Twin Towers Hotel & Convention Center
Orlando, Florida

Welcome Reception — April 20, 1993

3-Day Short Course and Workshop IMAGE ANALYSIS AND MEASUREMENT

May, 1993

North Carolina State University
Raleigh, North Carolina

Organized by John C. Russ, North Carolina State University

For additional information contact: Ms. Cindy Allen, Office of Continuing Education
North Carolina State University, Raleigh, NC 27695 • Phone (919) 515-2261

MSA PRE-MEETING WORKSHOP 1993

July 31, 8:00 am - August 1, 4:00 pm

Miami University
Oxford, Ohio

A hands-on practical experience in cryofixation & freeze-substitution

Organized by A. Allenspach, Miami University — (513) 529-3100
and M.V. Parthasarathy, Cornell University — (607) 255-1734

Invitees: John Kiss, Kent McDonald, and Martin Muller

Workshop Fees: MSA members \$225; other \$250 (to be paid before July 1, 1993)

Maximum number of participants: 20

FALL MEETING OF TSEM

October, 1993

Galveston, Texas

Details To Be Given Later

1993 SEM SPECIALS

SCINTILLATORS

\$81.00 MOST SEM'S

FILAMENTS

ISI - \$199/ BOX OF 10

Etec - \$249

AMRay - \$225

VACUUM FORELINE TRAPS

*WITH REPLACEABLE ELEMENT
FROM \$249*

SILVER ADHESIVE PRODUCTS

SILVER PAINT - 15 GRAMS \$14.95

SILVER PEN \$14.95

SILVER EPOXY \$21.95

SPECIMEN MOUNTS

CAMBRIDGE 1/2" dia., 1/8" peg 22 ¢ each

AMRay 1/2" dia., 1/8" peg 32 ¢ each

JEOL 3/8" dia. x 3/8" high 24 ¢ each

"Many more items - Custom Machining - Glassblowing"

M.E. Taylor Engineering Inc.
21604 Gentry Lane
Brookeville, MD. 20833

FAX: 301-774-6711

PH: 301-774-6246

NOW ON-LINE

The EMSA Electronic Bulletin Board System

1-800-627-EMSA

1-800-627-3672

Yes, finally it's here! The EMSA Electronic Bulletin Board System is off the backburners and is ready to be used and hopefully not abused. Interested? Just dust off your PC's, MAC's, or whichever system you prefer, get hold of a telecommunications program and a modem and log into the EMSA BBS. Set your modem for the following:

Modem Parameters: 1200 or 2400 Baud

8 Data Bits

1 Stop Bit

No Parity

Charges:

**None, EMSA is currently
sponsoring an 800 number**

Lines/Times:

One Line, Operating 24 Hours Per Day

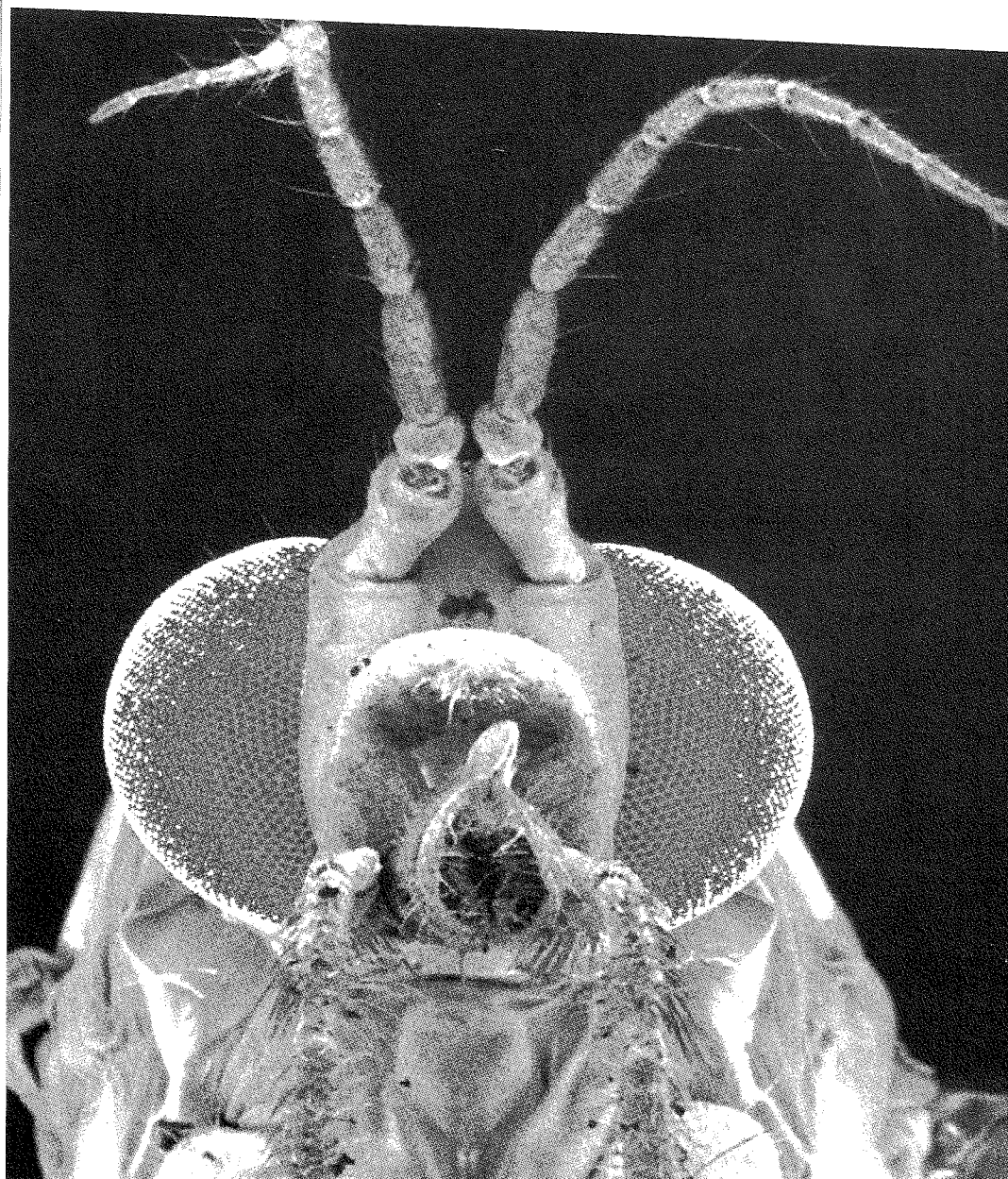
30 Minutes/User/Day

1-800-627-3672

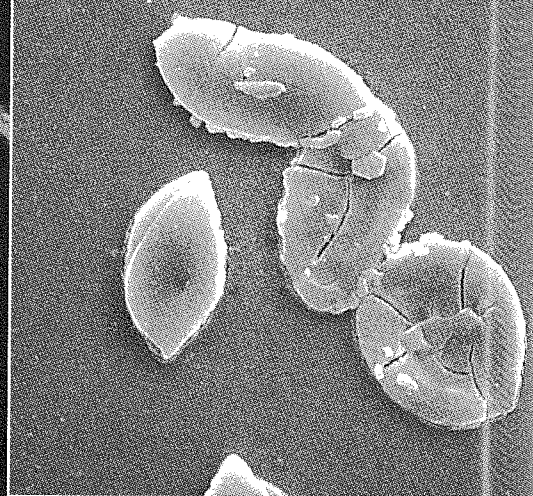
Upon login to the system, you will be prompted/asked several questions (and please use your real name!). Next you will be shown several notices and then set free to explore. A preliminary users manual is even available on-line. Electron Mail, Discussion Forums, EMSA notices/reports, Bulletin Articles, Meeting and Program information, special LAS areas and more will all be available in due course. If you want more details of have problems then stop by the Computer Workshop at the San Jose Meeting and we will try to help. Feel free to pass this information onto your colleagues as appropriate. See you in San Jose!

Nestor Zaluzec: EMSA BBS SysOp • Ron Anderson: EMSA BBS Chairman

TOPCON Scanning Electron Microscopes bring together 60 years of expertise in optics, mechanics and electronics to help science see nature eye-to-eye,



Blood cell responsible for sickle cell anemia.



to help medicine cure the incurable.

Integrated circuit with multiple film layers.



to make sure chipmaking processes stack up.

Take a closer look at Topcon.

Everywhere you look, you see TOPCON.

No other name appears on as many ophthalmic, medical and survey instruments worldwide. In every market we enter, TOPCON provides unique, differentiated products that allow users to do their best work. We are

number one in most of those markets, and that makes TOPCON the name to watch now in scanning electron microscopy.

Before blindly buying your next SEM, shouldn't you look into TOPCON first? For product availability and specifications, call us at 800-538-6850. Or write: TOPCON Technologies,

6940 Koll Center Parkway, Pleasanton, CA 94566 USA.

 **TOPCON**
TOPCON TECHNOLOGIES
INCORPORATED

ADVERTISERS INDEX

Advertiser	Page Located	Advertiser	Page Located
Barry Scientific, Inc.	30	Hitachi Scientific Instruments	39
Delaware Diamond Knives, Inc.	47	JEOL U.S.A., Inc.	28
Denka	46	Micro Engineering, Inc.	13 & 38
Denton Vacuum, Inc.	44	Oxford Instruments	2
Diatome U.S.	16 & 40	Ted Pella, Inc.	24
Electron Microscopy Sciences	10 & 32	Philips Electronic Instruments	14
EM Corp.	6	Structure Probe, Inc.	18
EMITECH U.S.A., Inc.	12	M.E. Taylor Engineering, Inc.	20
FEI Co.	8	TOPCON Technologies, Inc.	22
GATAN, Inc.	4	Carl Zeiss, Inc.	9

EDITORIAL POLICY

LETTERS TO THE EDITOR

Letters to the editor are printed as they are received in the order of their arrival. These letters reflect the opinion of the individual TSEM member and do not necessarily reflect the opinions of the editor or the society. The content of the letters should be concerned with the philosophical or operational aspects of the TSEM, the Journal and its contents, academic or national policies as they apply to TSEM and/or its members and electron microscopy in general. Editorial privilege may be evoked to insure that the LETTERS SECTION will neither be used as a political forum nor violate the memberships' trust.

ELECTRON MICROGRAPHS AND COVER PHOTOS

Micrographs submitted for cover photos should be marked as such. The choice of photographs will be made by the editor. Photograph receipt and/or dispensation will not be acknowledged. Photographs will not be returned. Electron micrographs to be used for cover photos and text fillers are welcome and should be selected with some attention to aesthetic appeal as well as excellence both in technique and in scientific information content.

REGIONAL NEWS

News items should be submitted through the regional editor in your area and made to conform to the standard format used by the regional news section. Regional contributions should be sent to the Regional News Editor. Editorial privilege may be executed for the sake of brevity or to preserve the philosophical nature of the TSEM Journal.

The JOB OPPORTUNITIES section will be comprised of a "Jobs Available" and a "Jobs Wanted" sub-section.

Anonymity of individuals listing in the Jobs Wanted or Jobs Available sub-sections may be maintained by correspondence routed through the Regional News Editor's office.

TECHNICAL SECTION

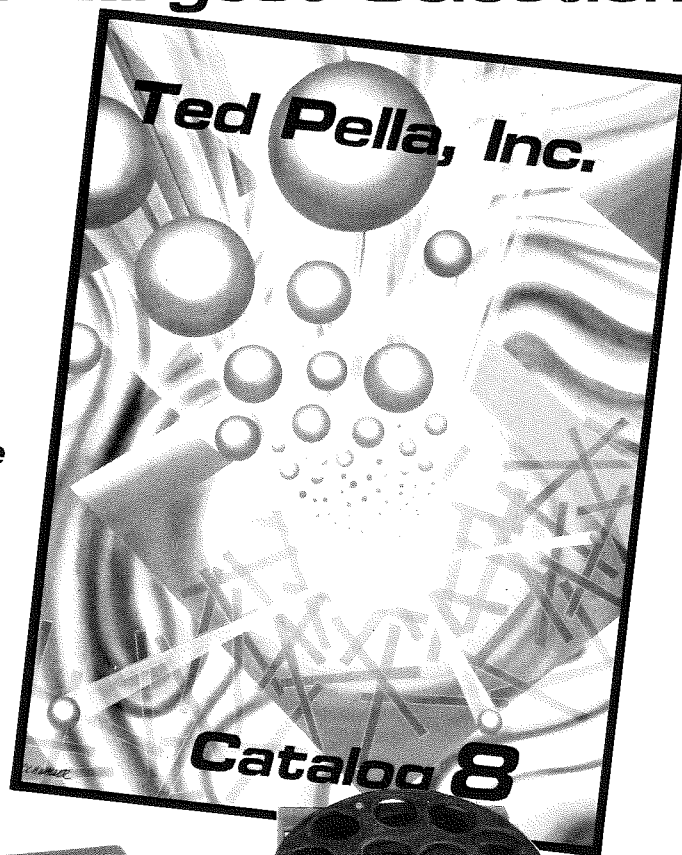
The Technical Section will publish TECHNIQUES PAPERS, HELPFUL HINTS, and JOB OPPORTUNITIES. The TECHNICAL PAPERS will describe new or improved methods for existing techniques and give examples of the results obtained with methods. The format of the Technique Papers will be the same as that used for regular research reports. HELPFUL HINTS will be in the form of a brief report with an accompanying illustration, if required for clarity. Helpful Hints should embody techniques which will improve or expedite processes and/or procedures used in EM.

PUBLICATION PRIVILEGES

The right to publish in the TSEMJ is restricted to TSEM members or to those whose membership is pending. A membership application form can usually be found in each issue of the TSEMJ. Membership dues are as follows: student \$2.00; regular members \$15.00; Corporate members \$75.00. Individuals who belong to TSEM by virtue of a corporate membership are invited to participate in Journal submissions as are our regular or student members. However, papers of a commercial nature, either stated or implied, will not be accepted for publication as a Research Report or Techniques Paper. Such papers may be acceptable as advertising copy.

Searching for EM Supplies? Catalog 8 Has the Largest Selection.

- Immuno Gold Conjugates
- Photo Supplies
- UV Cryo Chamber
- Vibrating Microtomes
- Grids & Tweezers
- SEM Specimen Mounts
- Graphic Transfer Lettering
- Filaments & Apertures
- Calibration Aids
- Trimming Stands ...And Much More



Request Your Copy Today.



Ted Pella, Inc.

The Electron Microscopy Supply Center

P.O.Box 492477, Redding CA 96049-2477

916-243-2200, 1-800-237-3526 (USA), 1-800-637-3526 (CA), 1-800-243-7765 (Canada), FAX: 916-243-3761

TSEM STUDENT COMPETITION

CASH • TRAVEL • RECOGNITION

ELIGIBILITY:

Competition is open to all student members of T.S.E.M. who are actively seeking a degree at an accredited institution. The term student member will also include those students with a membership application pending. To be eligible to compete, all competition requirements must be fulfilled by the designated deadlines given in the first call for papers preceding the Fall meeting. In addition, to be considered for the top award you must, (1) be a student at the time of the next EMSA meeting, (2) apply for a Presidential Student Award, and (3) present your paper at that meeting.

REQUIREMENTS:

You must be the sole author, personally present your paper from the platform, and submit a student competition application signed by a regular T.S.E.M. member, if possible your supervising professor. Two abstracts must be submitted by the designated deadlines; a regular T.S.E.M. abstract following normal procedures submitted to the current *Journal* editor, and an EMSA style two page abstract with an application for student travel submitted to the current secretary. Since it is assumed that your professor has supervised your work and others may have contributed in various ways, you must acknowledge these contributions on your application as well as in your platform presentation.

SPECIAL ABSTRACT FORMAT

1. The paper must be two pages each 8½" by 11". Margins should be 1" top and bottom and ¾" left to right. Text should be 12 characters per inch IBM LETTER GOTHIC or 11 point TIMES ROMAN with 12 point spacing each font at 6 lines per vertical inch.
2. The first page will have text only. Title on first line in all capitals except chemical symbols, single spaced if more than one line is needed. Leave one line of space; then your name and address skipping one line between each. Leave one line blank and start text with no indentions and skip one line between paragraphs. Group all references at the end on the text before illustrations.
3. Page two will include pictures and text. Micrographs should be numbered, have an appropriate scale marker, and be trimmed to form a rectangle with no gaps. Figure captions should follow the micrographs and come last.
4. Examples and additional guidelines may be found by consulting an EMSA call for papers.

AWARDS:

Up to 3 awards (0-3) may be given at each Fall meeting. These awards may be cash or prizes as determined by the Executive Council. The top award that can be given is substantial support towards competing in EMSA's Presidential Student Award program. This award can only be given if you meet EMSA qualifications and compete at the next EMSA meeting.

JUDGING:

Judging will be by a panel of regular T.S.E.M. members. You will be judged 50% on the quality of your special abstract and 50% on the quality of your presentation, including how well you answer questions from the audience. The regular abstract you submit for publication in the *Journal* will not be judged. Because of additional demands of disclosure each entrant will be given an additional 5 minutes of podium time.



TSEM STUDENT COMPETITION APPLICATION

Student's Name: _____

Mailing Address: _____

Phone: _____

University: _____

Department: _____ Supervising Professor: _____

Degree Program: _____ Anticipated Date of Degree: _____

Title of Paper: _____

Contributions from Others: _____

Do you wish to be considered for travel support to the next EMSA meeting?
By answering "YES", you agree to meet EMSA guidelines pertaining to the
Presidential Student Award program.

YES ____ NO ____

I certify that the work being reported is my own.

Student's Signature _____

I certify that the work being reported is that of the student.

Professor's Signature _____

Information for Authors

GENERAL INFORMATION

PURPOSE: The goal of the TSEM Journal is to inform members of the society and the Journal's readers of significant advances in electron microscopy, research, education, and technology. Original articles on any aspect of electron microscopy are invited for publication. Guidelines for submission of articles are given below. The views expressed in the articles, editorials and letters represent the opinions of the author(s) and do not reflect the official policy of the institution with which the author is affiliated or the Texas Society for Electron Microscopy. Acceptance by this Journal of advertisements for products or services does not imply endorsement. Manuscripts and related correspondence should be addressed to Louis H. Bragg, Editor, TEXAS SOCIETY FOR ELECTRON MICROSCOPY JOURNAL, Department of Biology, The University of Texas at Arlington, Box 19498, Arlington, Texas 76019.

GUIDELINES: Manuscripts written in English will be considered for publication in the form of original articles, historical and current reviews, case reports and descriptions of new and innovative EM techniques. It is understood that the submitted papers will not have been previously published. Accepted manuscripts become property of the TEXAS SOCIETY FOR ELECTRON MICROSCOPY JOURNAL and may not be published elsewhere without written consent of the Editor. The author should retain one complete copy of the manuscript. The JOURNAL is not responsible for manuscripts lost in the mail.

PAGE PROOFS/REPRINTS: The editor will be responsible for proof-reading the type-set article. Reprints may be ordered from the printer.

MANUSCRIPT PREPARATION: Manuscripts should conform with the following guidelines:

FORMAT: Submit an original and two copies of the entire manuscript, typed, double-spaced, on 8½ x 11 white paper, leaving ample margins. Number each page and identify the article by placing, at the top left of the page, a shortened form of the title, followed by the last name of the first author.

TITLE PAGE: Include:

- Full title of the article
- Initials and last names of all authors
- Current positions of each author (department, institution, city)
- Full name, telephone number and address of the author to whom reprint requests are to be sent.

SECTIONS: The text of each original article and technical report should be divided into four major sections entitled INTRODUCTION; METHODS AND MATERIALS; RESULTS; AND DISCUSSION.

Historical and current reviews and case reports do not need to be divided into the aforementioned sections.

ABSTRACT: Summarize the article in no more than 150 words. This takes the place of a final summary paragraph.

REFERENCES to other work should be consecutively numbered in the text using parentheses and listed at the end, as in the following examples:

- (1) A. Glauret, Practical Methods in Electron Microscopy. Vol. 2 (North-Holland. Amsterdam, 1974) 82-88.
- (2) P.S. Baur, Jr., G.F. Barratt, G.M. Brown and D.H. Parks. Ultrastructural Evidence for the Presence of "Fibroblasts" and "myofibroblasts" in Wound Healing Tissues. J. of Trauma. 19 (1979) 774-756.
- (3) D. Gabor. Information Theory in Electron Microscopy, in: Quantitative Electron Microscopy. Eds. G.F. Bahr and E. Zeitler (Williams and Wilkins, Baltimore, 1956) 63-68.

(NOTE: Authors are responsible for the accuracy of references.)

TABLES:

- Type double-spaced each table on a separate sheet.
- Number in order in which they are referred to in the text.

ILLUSTRATIONS:

- Submit three complete sets of illustrations. Copy machine reproductions of photographs will not be accepted. Indicate which set is the original photograph or illustration.
- Number the figures in the order in which they are referred to in the text.
- For black and white illustrations, submit sharply focused, glossy prints, or line drawings, 1.5 times larger than they are to appear in print (1/4 or 1/2 page). Scale should be drawn on the photograph itself, not below.
- For color illustrations, if needed, submit positive 35-mm color transparencies (not prints) for the original (prints may be used for the two copies). Authors will bear the entire cost of color reproductions.
- Identify all illustrations (author, title of paper, and number) by a gummed label on the back of each. Do not mount the illustrations, write on the back of them, clip them, or staple them.
- Illustrations taken from other publications require reprint permission and must be submitted in the form described above.

NOMENCLATURE AND ABBREVIATIONS: Journal abbreviations used should be those listed by the "Index Medicus." Nomenclature abbreviations should be similarly standardized.

ACKNOWLEDGEMENTS should appear as a footnote which will appear at the top of the first page of the article.

JEM-1210: A TOTALLY NEW CONCEPT IN TEM DESIGN

Our new JEM-1210 is the first to incorporate modern 32 bit computer technology into a state-of-the-art TEM. The resulting instrument offers biologists and materials scientists new levels of microscope performance.

In the 1210, biologists and materials scientists will achieve greatly expanded productivity and ease-of-use:

The operator interface has been radically changed to simplify operation while maintaining or even improving performance. For

example, essential manual controls—e.g. focusing and brightness knobs—have been retained, but now most operating parameters are mouse controlled.

A motorized objective aperture and a motorized goniometer (5 axis) extend remote control even to contrast and specimen manipulation.

Alignment, focus and brightness are adjusted, automatically, throughout the full magnification range.

Optimum operating conditions for a given specimen—magnification, beam current, image wobbler, through focus and minimum dose—are easily stored and may be quickly recalled.

Images may be digitized, displayed, manipulated to enhance features of interest, and either stored on one of several magnetic media or printed using a high resolution graphics printer.

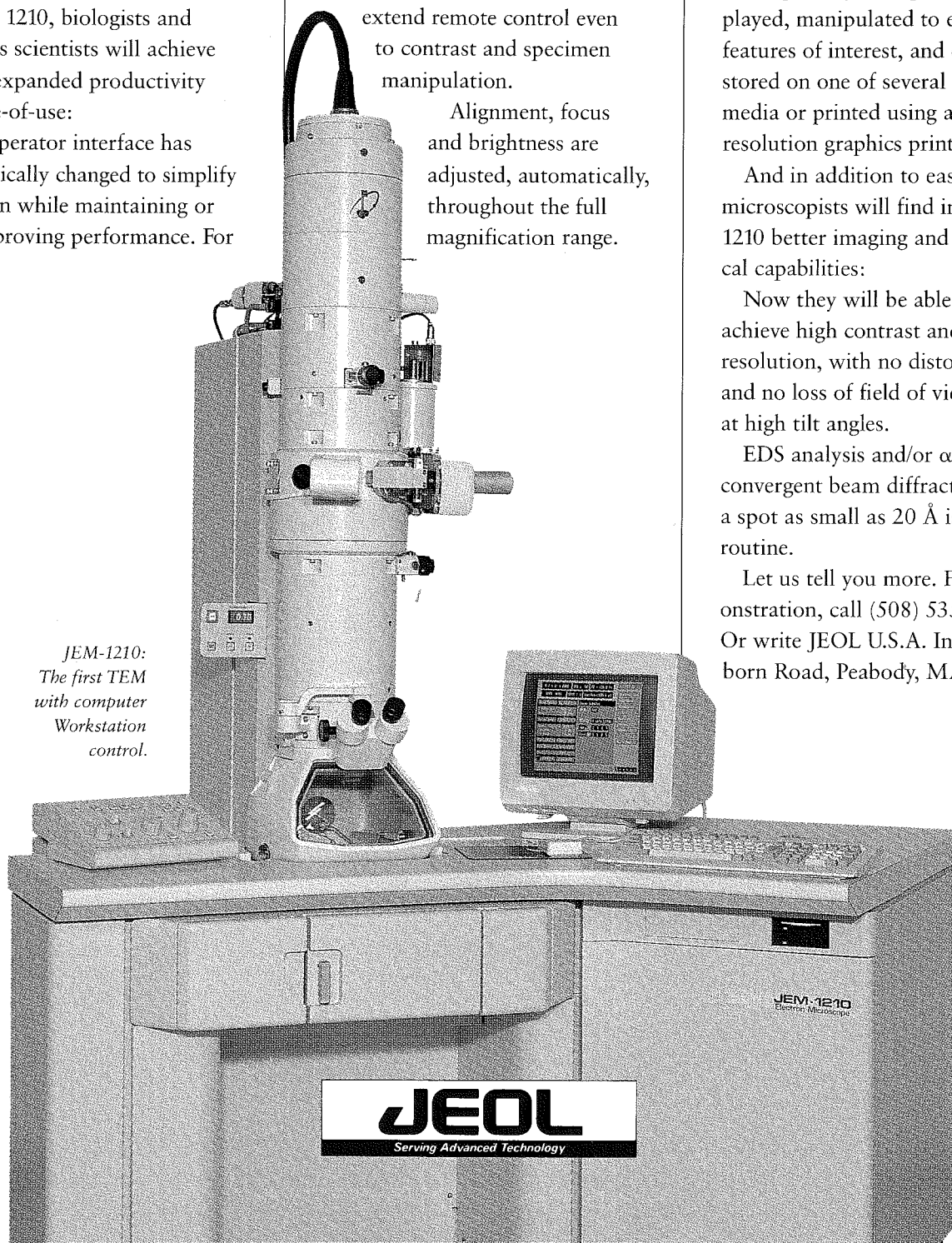
And in addition to ease-of-use, microscopists will find in the 1210 better imaging and analytical capabilities:

Now they will be able to achieve high contrast and high resolution, with no distortion and no loss of field of view, even at high tilt angles.

EDS analysis and/or α -selected convergent beam diffraction with a spot as small as 20 Å is now routine.

Let us tell you more. For a demonstration, call (508) 535-5900. Or write JEOL U.S.A. Inc., 11 Dearborn Road, Peabody, MA 01960.

*JEM-1210:
The first TEM
with computer
Workstation
control.*



JEOL
Serving Advanced Technology

MSA UNDERGRADUATE SCHOLARSHIP PROGRAM

Description and Eligibility Requirements

1. The Microscopy Society of America (MSA) has established a scholarship fund for undergraduate research. The purpose of the scholarship is to further educational and research potential in full time undergraduate students intent on pursuing electron microscopy as a career. Applications in all areas of electron microscopy will be accepted for review.
2. Scholarships will be awarded to full time undergraduate students. Maximum total dollar amount awarded each year will be \$10,000.00.
3. When possible at least one scholarship will be awarded to an under-represented minority applicant.
4. Preference will be given to those scholarship proposals which utilize a facility other than the one at which the student is currently enrolled.
5. Applicant must be a full time undergraduate student and a U.S. citizen or resident alien (green card required). Research programs for which scholarship funds are awarded must be carried out in a U.S. laboratory.
6. Awarded funds must be used within a designated time period not to exceed one year from the award date. Students are eligible to receive an award only once.
7. Applications must be received by 15 November of each year to be awarded by 1 March of the following year.
8. Complete application forms should include the following items:
 - A. Provided form with requested information.
 - B. A research proposal not to exceed 3 pages in length. The proposal should include a brief introduction, a short methods section, and itemized goals of the study.
 - C. A budget proposal detailing how awards will be utilized. If the proposed project will exceed the requested amount, a statement should be provided indicating sources of additional funding.
 - D. Two letters of reference from academic and/or industrial personnel familiar with the student's competence are required.
 - E. A letter from the laboratory supervisor where the proposed research will be performed indicating the applicant will be accepted in the laboratory to work on the proposed project. A laboratory must be designated in the proposal for funds to be awarded.
 - F. A curriculum vitae detailing previous education and/or experience in electron microscopy, and a brief statement of career goals.
9. Complete applications, reference letters, and a letter from the laboratory supervisor where the research will be performed should be sent to:

Dr. M.G. Burke
Westinghouse Science & Technology Center
1310 Beulah Road
Pittsburgh, PA 15235-5098
10. Following completion of the scholarship period a statement detailing status of the research project should be given to the MSA Council. Publications resulting from the scholarship should acknowledge the award. A reprint should be given to the MSA Education Committee Library when they become available.

BARRY SCIENTIFIC

Kimball Physics LaB₆ CATHODES with MULTI-THOUSAND HOUR LIFETIMES

EXTENDED LIFETIME / SUPERIOR QUALITY

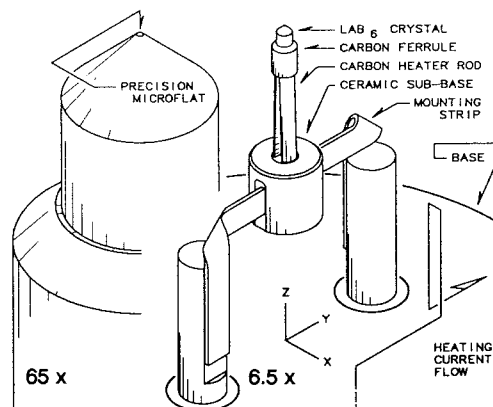
Thousands of Hours of Life with Clean Technique
100 μm of LaB₆ Available Surface Loss Guaranteed
Precision-Machined Stress-Free Carbon Mounting
Reduces Mounting Structure Failures

HIGH BRIGHTNESS / LOW ENERGY SPREAD

Available Brightness Above 1×10^6 Amp/cm² steradian
Oriented Single-Crystal - <100> Standard
Best Quality / High Purity Material
Accurate Microflats / Controlled Crossover Size

EXCEPTIONAL STABILITY / EASY OPERATION

Thermal / Chemical / Mechanical / Electrical Stability
High Over-Temperature Tolerance
Fits All Bases / Custom Units Available
Extensive Application Notes on Request



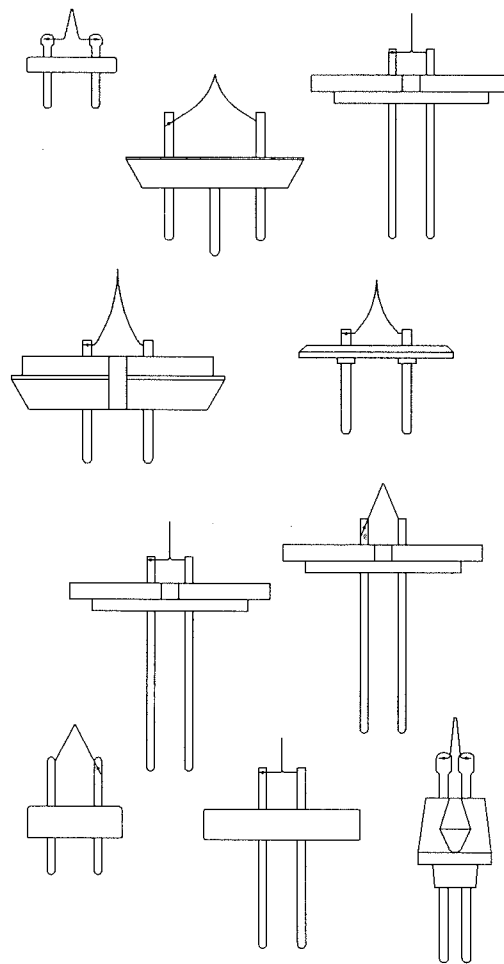
Model ES-423E Lanthanum Hexaboride Cathode:
6.5 x enlargement shows mounting structure.
65 x enlargement shows precision microflat emitting surface.

TUNGSTEN SEM/TEM FILAMENTS

BEST QUALITY / FULLY ANNEALED
PRECISION ALIGNED / STRAIN FREE
NEW PACKAGING / INDIVIDUALLY SECURED
FILAMENTS VISIBLE in Unopened Box
EASY-TO-USE HANDLING TOOL Included at No Charge

Kimball Physics tungsten filaments, new or rebuilt, are formed in high-precision fixtures using a special-alloy tungsten wire to minimize strain. Precise dimensioning and microscopic inspection are done to guarantee consistent quality and performance. All filaments are fully annealed.

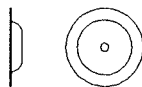
A new design in packaging allows a visual check of the filaments without opening the box. Also, if an open box is tipped over, the filaments will not fall out; each is held separately in place. With many instruments it is no longer necessary to touch the filaments. A filament handling tool is provided to transfer filaments directly into the instrument without manual contact. Reduce the danger of contamination and breakage; reduce the need for clean room gloves.



WEHNELT APERTURES

HIGH-QUALITY Ta APERTURES

Precision machined and polished tantalum apertures are supplied in three basic sizes: 500, 750, and 1000 micron diameter holes. Other hole sizes are available upon request.



Products Manufactured By

KIMBALL PHYSICS INC. KPI

Exclusive Retail Distributor for North America

BARRY SCIENTIFIC

P.O. Box 173, 58 Streeter Road, Fiskdale, Massachusetts 01518
Tel (508) 347-9855 1-800-348-2257 Fax (508) 347-8280

MSA UNDERGRADUATE SCHOLARSHIP APPLICATION

1. Name (Last, First, Middle): _____
2. Date of Birth: _____ 3. Social Security Number: _____
4. U.S. Citizen: ☐ Yes ☐ No Resident Visa: ☐ Yes ☐ No
5. Telephone Number (Area Code, Number, Extension): _____
6. Present Address: _____ Permanent Address: _____
- _____
- _____
- _____
7. University Affiliation: _____
8. Academic Status (Year Completed by Spring): ☐ Fr. ☐ So. ☐ Jr. ☐ Sr.
9. Title of Proposed Research: _____
10. Site where proposed research will be conducted: _____
- _____
11. Amount of funding requested (Maximum = \$2,500): _____
Include a detailed budget with sources of additional support for project on a separate page.
12. Name, title, and addresses of two academic or industrial personnel familiar with the student who will provide letters of reference. Applicant is responsible for having all letters sent before the application deadline. Letters must be received before application is considered for funding.
- _____
- _____
- _____
- _____
13. Name, title, and address of laboratory supervisor who will supervise the project and provide a letter of reference.
- _____
- _____
- _____
- _____
14. Optional Information: Racial/Ethnic Background: _____
(Note: When possible, at least one award will be given to an under-represented minority applicant.)
15. Give a brief statement concerning your chosen academic major, and your career objective.
- _____
- _____
- _____
16. I certify that I am a full time undergraduate student, and that all information provided in this application is true. Any false information may result in forfeiture of awarded funds.
- Applicant's Signature: _____ Date: _____

The Chemicals You Want The Quality and Value You Need

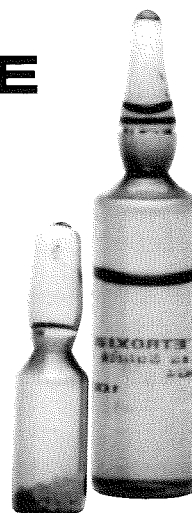
At Electron Microscopy Sciences, We've Built a Business on it!



GLUTARALDEHYDE

For over 20 years we have been manufacturing the highest purity Glutaraldehyde available on the market; free from polymers and other contaminants. Prior to filling each lot is tested and assayed to assure consistent purity. Only if the Glutaraldehyde passes our rigorous quality control tests will we ship it to you.

Our EM grade is available in 8%, 10%, 25%, 50%, and 70% in 2ml, 5ml, 10ml ampoules as well as 100ml bottles. Our Biological grade is available in 25%, and 50% in 450ml and 1 gallon containers.



OSMIUM TETROXIDE



Crystalline (99.95%) and Solution

Each glass ampoule is pre-scored, pre-cleaned, and heat sealed in a plastic bag - guaranteeing you a contaminant-free solution.

Our solution is available in standard concentrations of 2%, and 4%, in 2ml, 5ml, and 10ml ampoules.

Our crystalline is available in 6gm, 5gm, 4gm, 2gm, 1gm, 1/2gm, 1/4gm 1/10gm ampoules.

Quantity discounts available - please call for special pricing.

Here at Electron Microscopy Sciences we have perfected the manufacturing and filling of the highest quality chemicals meeting all of your microscopy needs. In addition to the chemicals that are listed in our catalog we accept all special orders. If you have special size requirements, concentrations or purity specifications,

Electron Microscopy Sciences is the source.

For a copy of our newest catalog of supplies, accessories, chemicals, and equipment covering the entire field of Microscopy call or write us today.

For the best results in your valued research, look for the name that is leading the way in the highest quality chemicals meeting all of your microscopy needs.

321 Morris Road • Box 251 • Fort Washington, PA 19034
Toll-free: 1-800-523-5874 • (215) 646-1566 • Fax: (215) 646-8931 Telex: 510-661-3280

**Electron
Microscopy
Sciences**

Abstracts

BIOLOGICAL SCIENCES

PLATFORM PRESENTATION — SPRING 1993

IMAGING PERISTOME MOVEMENT IN THE ENVIRONMENTAL SEM
J.C. Long, D.M.J. Mueller, Electron Microscopy Center, Dept. of Biology, Texas A&M University, College Station, Texas 77843-2257

The ElectroScan Environmental Scanning Electron Microscope is a powerful new tool in the study of spore dispersal mechanisms in bryophytes. The ESEM is similar in many ways to a conventional SEM, but with a few important differences. While the ESEM column remains at high vacuum (10^{-7} torr), the specimen chamber typically operates in a water vapor environment at low vacuum (1 to 20 torr). The secondary electron signal is generated by gaseous signal amplification and is collected by the Environmental Secondary Detector.

This new technology has significant implications regarding sample preparation and specimen requirements. First, there is no need to dry the sample, thus observations in the natural state are possible. Second, the water vapor dissipates charge build up, therefore a conductive coating is not necessary.

The ESEM lends itself directly to the understanding of spore dispersal in mosses, which involves the sensitivity of the peristome to changes in ambient moisture (hygroscopicity). Using conventional SEM techniques, it is not possible to view peristome movement directly, however, using the ESEM, the movements can be observed and recorded in real time.

In many mosses the peristome is an integral part of the spore release mechanism which regulates the release of spores from the capsule. Peristomes consist of the thickened portions of concentric rings of cell walls of contiguous layers of cells. These cell walls remain responsive to changes in ambient moisture and exhibit movement with these hygroscopic changes in such a way as to effect the release of spores (facilitating the release in some mosses, hindering the too rapid release of spores in others).

Peristome movements are recorded, in real time, on videotape with sufficient magnification and resolution to view, for the first time, the intricacies of movement as it reflects the variable composition and construction of the cell wall layers making up these structures.

INTERACTION OF *PSEUDOMONAS AERUGINOSA* WITH POLARIZED EPITHELIAL CELLS. LYNN D. GRAY and ALI O. AZGHANI*, Dept. of Cell Biology and Ev. Sci., *Dept. of Biochemistry, The University of Texas Health Center at Tyler, P.O. Box 2003, Tyler, TX 75710.

Pseudomonas aeruginosa, an opportunistic pathogen, causes a variety of illnesses including pneumonia, septicemia and chronic lung disease. The organism is a common cause of fatal nosocomial infections in immunosuppressed patients. Relatively little is known about how this bacterium attaches and invades host tissues and what roles its various secretory products play in pathogenicity. *In vivo* studies in our laboratories showed increased epithelial permeability in the lungs of guinea pigs that were treated with elastase (PE) from *P. aeruginosa*. The present experiments explore *in vitro* host-pathogen relationships between this organism and two types of epithelial cells. Confluent Madin-Darby Canine Kidney (MDCK) cells exhibit "tight" cell junctions (based on morphology and electrical resistance) and provide a homogeneous system to study the effects of PE and other bacterial products on epithelial cells. Primary, cultures of rat type II pneumocytes were used in some experiments because of our interest in pulmonary effects. Some monolayers were exposed to live bacteria and incubated at various time periods from 2 to 6 h. Others were treated with PBS, or PE (0.06 - 6u/ml) prior to application of live bacteria. Qualitative observations were made in a single-blind fashion. SEM of the MDCK cells revealed scattered eroded areas containing attached bacteria as early as 3 h post-exposure. Other pathologic features included alteration and reduction of microvilli and thick, mucoid secretions on affected cell surfaces. Bacteria were often found embedded in this material. Bacteria were consistently found in association with clumps of dividing cells, at cell borders and in eroded areas of the MDCK monolayers; they were located near cell junctions and in regions of lamellar body secretion in the rat type II cells. *P. aeruginosa* readily adhered to both types of epithelial cells and SEM was essential in assessing the number and location of attached bacteria. This work is supported by Grants from the American Heart Association (Texas Affiliate) and NIH/LBI #HL44473.

SOFT TISSUE SARCOMAS RESEMBLING PRIMARY BONE TUMORS.

Bruce Mackay, Alberto G. Ayala, Nelson G. Ordonez.
University of Texas M.D. Anderson Cancer Center, Houston, Texas.

Occasional malignant tumors arising in the soft tissues appear identical in microscopic sections to primary tumors of bone and cartilage. Extraskeletal osteosarcoma and chondrosarcoma are soft tissue neoplasms which closely resemble the corresponding bone tumors by light microscopy and at the ultrastructural level. Care must be taken to avoid mistaking myositis ossificans for an extraskeletal osteosarcoma. Electron microscopy has been of some value in the study of certain unusual soft tissue tumors within the extraskeletal category. A type of small round cell tumor originating in the soft tissues looks like Ewing's tumor of bone by light microscopy and is called soft tissue Ewing's sarcoma. The cells show the same range of appearances by electron microscopy. There has been much speculation on the cell of origin but it remains unknown. Chondrosarcoma subtypes comparable to those seen within bone occur in the soft tissues, and some extraskeletal myxoid chondrosarcomas contain large numbers of parallel microtubules within cisternae of the endoplasmic reticulum. Parachordoma is a rare soft tissue neoplasm that simulates extraskeletal myxoid chondrosarcoma by routine light microscopy but it possesses epithelial characteristics which are revealed by immunostaining and electron microscopy; its close resemblance to the sacro-coccygeal chordoma suggests that it is derived from ectopic notochord cells.

THE ROLE OF DIAGNOSTIC ELECTRON MICROSCOPY IN NEURO-ONCOLOGY: SOME OLD AND SOME NEW OBSERVATIONS WITH THERAPEUTIC IMPLICATIONS. Steven C. Bauserman, J.C. Stinson Laboratory for Electron Microscopy, Scott and White Clinic, Texas A&M University Health Science Center, Temple, Texas 76508

With the advent of immunohistochemistry the application of Transmission Electron Microscopy (TEM) in diagnostic pathology has diminished to some extent. In surgical pathology of the central nervous system, however, there is a persistent need for this adjunctive study in several specific clinical settings. In one particular type of tumor the application of TEM has identified an entity which has masqueraded in the past as **Oligodendroglioma** or **Clear Cell Ependymoma** and carries a much better prognosis. This is the newly recognized **CENTRAL NEUROCYTOMA** which usually projects into the ventricular system of the brain with a very good prognosis if surgically resected. Examples of this particular tumor are presented with their distinctive radiographic and pathologic as well as ultrastructural features. Additional applications including **Metastatic Neoplasm of Brain with unknown primary**; **Primitive Neuroectodermal Tumor (PNET)** of infancy and childhood; and some **Sarcomatous lesions of the brain and its coverings** are presented with discussion of the adjunctive role of TEM in diagnosis.

A COMPARISON OF SIX GOLD PARTICLE SIZES ON LABELING DENSITY USING GOLD CONJUGATED GOAT ANTI-RABBIT IgG SECONDARY ANTIBODIES. RICK GIBERSON, Ted Pella, Inc., Redding, CA 96049-2477

B-lactoglobulin granules found in processed cheese were indirectly labeled with six different sized gold conjugated goat anti-rabbit IgG whole antibodies. The effects of gold particle size on labeling density (particles/ μm^2) were determined for 1, 5, 10, 15, 20, 30nm gold particles. A relationship between gold particle size and labeling density was found with IgG conjugated gold. Immunogold labeling with the ultra small (1nm) IgG conjugated gold, followed by silver enhancement, demonstrated the lowest labeling density.



IMMUNOGOLD LOCALIZATION OF PECTIN EPITOPES IN THE CELL WALL OF BORON-DEFICIENT CALLI OF *LYCOPERSICON ESCULENTUM* MILL. AND *OXALIS DILLENII* JACQ. CAMELIA G.A. MAIER*, DON W. SMITH AND DAVID C. GARRETT, Biological Sciences Dept., University of North Texas, Denton, TX 76203.

Cell wall structure and composition are known to be affected by boron deficiency. In this study, monoclonal antibodies recognizing methyl-esterified (JIM 7) and un-esterified (JIM 5) epitopes of pectin have been used to locate these epitopes by immunogold electron microscopy in the cell walls of boron-deficient calli from tomato and oxalis.

Both antibodies labelled the cell wall of calli under study. The epitope containing methyl-esterified pectin was localized evenly throughout the cell walls of both boron-deficient and boron-sufficient (control) calli. The un-esterified epitope of pectin was located toward the inner surface of the primary cell walls, in the middle lamella, and abundantly toward the outer surface at intercellular spaces of boron-sufficient calli. In boron-deficient calli of both tomato and oxalis, the un-esterified epitopes of pectin did not have a specific location in thin cell walls. In thicker cell walls, these epitopes were found toward the outer surface but not in the middle lamella. These results indicate differences in location on un-esterified pectin epitopes in cell walls of boron-deficient and boron-sufficient tomato and oxalis calli.

BIOLOGICAL SCIENCES

POSTER PRESENTATION — SPRING 1993

SYNAPTIC ORGANIZATION OF DOPAMINERGIC AMACRINE CELLS IN THE LARVAL TIGER SALAMANDER RETINA. P.A. Glazebrook and C.B. Watt. Alice R. McPherson Laboratory of Retina Research, Center for Biotechnology, Baylor College of Medicine, The Woodlands, TX 77380.

Immunocytochemistry of tyrosine hydroxylase (TH) was used to visualize tiger salamander dopaminergic amacrine cells and determine their basic synaptic interaction. The avidin-biotin immunoperoxidase method was used to immunostain TH immunoreactive cells in vibratome-prepared sections that were routinely processed for ultrastructural examination.

A total of 168 synapses were observed that involved tyrosine hydroxylase-like immunoreactive amacrine cell processes. A large percentage (79.8%) of these synaptic arrangements were found in sublayer 1 of the inner plexiform layer, while substantially lower percentages were observed in sublayers 3 (9.5%) and 5 (10.7%). They served as pre and postsynaptic elements 63.1% and 36.9% of the time, respectively. Tyrosine hydroxylase-like immunoreactive amacrine cell processes were presynaptic to amacrine cell processes (36.9% of total synaptic involvement) and processes that lack synaptic vesicles and whose origin remains uncertain (26.2%). They received synaptic input primarily from amacrine cell processes (31.0%). Tyrosine hydroxylase-like immunoreactive amacrine cell processes also received a few ribbon synapses from bipolar cells (5.9%). Each of these synaptic relationships were observed in each of sublayers 1, 3 and 5 of the inner plexiform layer, with the majority of each arrangement being found in sublayer 1. Supported by grants from the NIH (EY05622) and the Retina Research Foundation (Houston).

SEM EVALUATION OF END-ROUNDING OF TOOTHBRUSH BRISTLES. NANCY K.R. SMITH*, DEBRA D. SMITH, AND H. RALPH RAWLS, *Dept. Cellular and Structural Biology, Dept. Restorative Dentistry Division of Biomaterials, University of Texas Health Science Center at San Antonio TX 78284.

Toothbrushes with non-rounded bristles can cause significant tissue trauma. We used scanning electron microscopy (SEM) to examine and evaluate adult-size toothbrushes for their potential to cause tissue damage. End-roundness was measured on 15-30 specimens each of a non-end-rounded brush and eight American-made brushes. Brush bases were cut with a Jim Dandy saw into 5-tuft sections which were then cleaned and superglued onto Al specimen stubs. The plastic bases and the bases of the nylon tufts were painted with colloidal graphite. Specimens were sputter-coated with Au-Pd. Five tufts from each brush were photographed at 50X in a JEOL JSM-35 SEM at 5kV, WD39mm. In each tuft 5 bristles were randomly selected and traced with a digitizer pad for calculation of the shape factor (SF), where $SF = \sqrt{\text{sq. root of tip area/perimeter}}$. The closer a

bristle tip is to being hemispherical ($SF = .28$), the higher its SF; the more sharp-edged, the lower. Brushes were ranked according to the % of bristles that exceeded a threshold SF value of .27, using Chi-squared analysis of variance. The ranking was: Oral-B P35 (73%) = Sensodyne (72%) = J&J Reach (70%) \geq POH (66%) = Colgate Plus (64%) = Pycopay (63%) \geq J&J Prevent (61%) = Butler GUM (58%) \geq no end-rounding (52%). The proportion of bristles with the greatest potential for harm ($SF < 0.25$, i.e., right-angled or sharper edges) were: Oral-B P35 = 1%, Sensodyne = 1%, Reach = 2%, Pycopay = 2%, POH = 3%, Colgate Plus = 5%, J&J Prevent = 5%, Butler GUM = 10%, no end-rounding = 4%. It was concluded, for the brushes examined, that Oral-B P35 had the least potential for tissue damage, Butler GUM had the most, and the others were intermediate.

SYNAPTIC INPUT TO GABAergic GANGLION CELLS IN THE INNER PLEXIFORM LAYER OF RABBIT RETINA. P.J.G. Neill and K.R. Fry. Alice R. McPherson Laboratory of Retina Research. The Center for Biotechnology, Baylor College of Medicine, The Woodlands, TX 77381.

Studies in this laboratory have identified a population of ganglion cells in the rabbit retina which utilize γ -aminobutyric acid (GABA) as a neurotransmitter. There is evidence to suggest that these cells may project to the area of the hypothalamus which is involved in circadian rhythm entrainment. Therefore, an understanding of the types of information carried by these cells and their pattern of synaptic connectivity in the retina is of great interest. Ganglion cells were first identified by pre-embedding immunoperoxidase labelling using a ganglion cell specific monoclonal antibody (AB5) developed in this laboratory. Post-embedding gold labeling was performed using an anti-GABA polyclonal antibody. Six percent of AB5 labelled ganglion cell processes also labelled with GABA, correlating well with previous light microscopy studies which indicated that 5-7% of ganglion cells were GABAergic. GABAergic ganglion cell processes received input from bipolar cell processes in the region of the inner plexiform layer (IPL) generally thought to mediate the "off" signals. The other primary synaptic input to the GABAergic ganglion cells was from GABAergic amacrine cell processes in sublaminae 1, 3, and 5 of the IPL; the frequency of synaptic input increased proportionately with closer proximity to a GABAergic ganglion cell body. These results indicate that ganglion cell processes receive direct input from the "off" visual pathway with modulation from GABAergic amacrine cells occurring throughout the IPL. Studies have been started to examine the distribution of other neurotransmitter-specific types of input to the GABAergic ganglion cell pathway. (Supported by NIH EYO6469 and the Retina Research Foundation)

A DETERMINATION OF ADULT FEEDING HOST RANGE AND LONG DISTANCE MIGRATION PATTERNS OF CORN EARWORM, CABBAGE LOOPER, AND CELERY LOOPER MOTHS UTILIZING SCANNING ELECTRON MICROSCOPY. P.D. Lingren¹, V.M. Bryant, Jr.², J.R. Raulston¹, M.W. Pendleton², J. Westbrook¹, & R.E. Murry². ¹USDA-ARS-CIPMRU, Rt.5, Box 808, College Station, TX 77845, ²Dept. of Anthropology, Texas A&M University, College Station, TX 77843.

Several species of moths feed on nectar-producing plants. Adult moths contact pollen grains during feeding activities and these grains often adhere to moth bodies. Identification of the pollen grains adhering to 400 moths captured from southern Texas and southern Oklahoma was made utilizing scanning electron microscopy. On the basis of these identifications, host plant feeding range and possible migratory activities of corn earworm, *Helicoverpa zea* (Boddie), cabbage looper, *Trichoplusia ni* (Hubner), and celery looper, *Anagrapha falcifera* (Kirby) moths were determined. A wide range of pollen grain taxa such as *Citrus*, *Salix*, *Quercus*, and *Pithecollobium* was observed on these moths. Eight percent of the *H. zea* moths captured in southern Oklahoma and 30 percent of those captured in southern Texas had *Citrus* pollen adhering to them. Associated weather systems and atmospheric trajectories suggest that captured moths associated with *Citrus* pollen had been conveyed by these forces at least 700 km northward because *Citrus* is not native to Oklahoma.

MATERIALS SCIENCES

PLATFORM PRESENTATION — SPRING 1993

COMPARATIVE OBSERVATIONS OF CARBIDE PRECIPITATION MORPHOLOGY ASSOCIATED WITH COHERENT TWIN BOUNDARIES AND GRAIN BOUNDARIES IN TYPE 304 STAINLESS STEEL: R. J. ROMERO, E. A. TRILLO, A. H. ADVANI, L. E. MURR and W. W. FISHER, Department of Metallurgical and Materials Engineering, The University of Texas at El Paso, El Paso, TX 79968.

When 304 and 316 stainless steels are aged at increasing times, carbides appear to occur first on high energy grain boundaries then non-coherent twin boundaries, and finally on coherent twin boundaries when selectively etched and observed by light microscopy. In order to study carbide nucleation, it was assumed that the coherent twin boundary would be invariant in structure and energy and would therefore be an ideal situation. However efforts to document carbide precipitation on coherent twin boundaries has been inconclusive because the preponderance of carbides apparently nucleate and grow from non-coherent steps on the boundary. In addition, the precipitates tend to grow parallel to the coherent boundary strongly suggesting coincidence with {111}. By comparison, carbides growing from grain boundaries also appear to nucleate prominently on steps or ledges in the boundary, and they grow out from the boundary coincident or partially coincident, with {111} planes in one principal grain. We have made numerous comparisons of these growth features using transmission electron microscopy. Supported by NSF-RIMI Grant HRD 9105065 and EPA Cooperative Agreement CR-81 8196-01-0.

THE EFFECT OF NICKEL ON Sn-Cu INTERMETALLIC GROWTH, Y. WU, J.A. SEES, C. POURAGHABGHER, E.G. JACOBS AND R.F. PINIZZOTTO, Center for Materials Characterization, University of North Texas, Denton, TX 76203-5308.

The intermetallic compounds Cu_6Sn_5 and Cu_3Sn form and grow at the Sn-Pb solder/copper substrate interface during soldering and system use. The addition of Ni particles to eutectic Sn-Pb solder drastically increases the activation energies of formation for both Cu_6Sn_5 and Cu_3Sn . To obtain direct information about the mechanisms of Cu-Sn intermetallic formation and the effect of Ni additions on intermetallic growth at the solder/substrate interface, Cu/Sn/Ni thin film samples were observed in real time using a hot stage in the TEM.

Standard TEM grids with thin amorphous carbon were used as the substrate for deposition of thin layers of Cu, Sn, and Ni by evaporation. In one set of samples, a continuous 500 Å thick layer of Cu was evaporated onto the substrate. A 500 Å layer of Sn was then deposited using a 50 mesh TEM grid as a shadow mask. After shifting the mask grid, 100 Å of Ni was deposited. This configuration results in Ni on top of part of the isolated square Sn-islands on Cu substrates. At 250°C, intermetallic formation begins at the Sn/Cu boundary and progresses laterally from the Sn into the Cu in some areas; in other areas, there is no intermetallic growth even after annealing at 250°C for 30 minutes and 200°C for 24 hours. XEDS spectra prove that in the areas with Cu-Sn intermetallic growth, there is no Ni on top of the Sn or at the interface; in the areas without intermetallic growth, there is Ni at the Cu/Sn interface, and there is a Ni concentration gradient from the Cu/Sn interface to the Sn area. This proves that Ni acts as a barrier which prevents Sn from diffusing into the Cu substrate and results in the drastic increase of the activation energies for the formation of both Cu_6Sn_5 and Cu_3Sn in Ni composite solder.

METALLURGICAL TECHNIQUES FOR ESTIMATING MICROMETEOROID IMPACT VELOCITIES FROM CRATER GEOMETRIES: JESUS RIVAS and L. E. MURR, Department of Metallurgical and Materials Engineering, The University of Texas at El Paso, El Paso, TX 79968.

We have observed numerous hypervelocity micrometeoroid impact craters in aluminum and stainless steel samples exposed in low-Earth orbit on the NASA Long Duration Exposure Facility (LDEF) satellite using normal-view scanning electron microscopy (SEM). In an effort to examine the crater wall structure and prospects for impact-generated shock-wave induced defects below the crater base, a technique was developed to produce cross-sections and slices (~20 µm) through larger-sized craters (>0.5 mm diameter) to allow SEM views normal to the crater wall, or transmission electron microscopy (TEM) of thin regions below (or beyond) the crater wall by argon ion milling of regions about 1 crater diameter from the wall. On comparing these section views with computer simulations in half-section space as a function of impact velocity, it was observed that features of the crater geometry such as the ejecta rim width and the height of the ejecta rim also varied with crater depth and diameter. It now appears that ratios of crater ejecta rim width to crater diameter, and ejecta rim height to crater diameter (or depth), may provide some estimates of the impact velocity, and we will present some examples of estimates based on observations of cross-sections as well as normal crater views. Work supported by NASA-Johnson Space Center Grant NAG 9-481.

THE OBSERVATION OF SILICON NANOCRYSTALS IN SILOXENE, R.F. PINIZZOTTO, H. YANG and J.M. PEREZ, Center for Materials Characterization and Physics Department, University of North Texas, Denton, Texas, 76203-5308 and J.L. COFFER, Department of Chemistry, Texas Christian University, Fort Worth, Texas, 76129.

Observations of the visible photoluminescence of many silicon-based materials have recently been reported. There is strong interest in understanding the basic luminescence mechanisms both from a fundamental physics perspective and for optoelectronic applications. We have used high resolution transmission electron microscopy to examine unannealed siloxene and have observed the presence of silicon nanocrystals with dimensions on the order of a few nanometers embedded in the material. This observation is additional strong evidence that the photoluminescence properties of Si-based materials are due to quantum confinement effects. The observations stress the underlying importance of HREM evaluation of photoluminescent silicon-based materials to fully understand the nature of this phenomenon.

DIFFUSION KINETICS OF INTERMETALLIC COMPOUND FORMATION IN COMPOSITE SOLDER. J.A. Sees, Y. Wu, J.L. Marshall, R.F. Pinizzotto, Center for Materials Characterization, University of North Texas, Denton, TX 76203-5308

The Sn/Pb eutectic alloy system is the most widely used joining material in the electronics industry. In this application, the solder provides the electrical and mechanical interconnection between integrated circuits on a printed circuit board. In an effort to improve its mechanical integrity, metallic or intermetallic particles have been added to eutectic Sn/Pb solder to form composite solder. The growth and morphology of the two intermetallic phases (Cu_6Sn_5 and Cu_3Sn) that form between a Cu substrate and Sn/Pb solder were studied under different aging and annealing conditions using scanning electron microscopy and X-ray energy dispersive spectroscopy. Activation energies for formation of these phases were determined for eutectic and four types of composite solder (20 wt.% Cu_6Sn_5 , 20 wt.% Cu_3Sn , 7.6 wt.% Cu, and 4.5 wt.% Ni). Cu-containing particles increased the activation energy of Cu_6Sn_5 formation and decreased the activation energy of Cu_3Sn formation. Ni additions dramatically increased both activation energies.



EFFECT OF CARBON CONTENT ON CARBIDE PRECIPITATION IN 304 STAINLESS STEEL: E. A. TRILLO, A. H. ADVANI, and L. E. MURR, and W. W. FISHER, Department of Metallurgical and Materials Engineering, The University of Texas at El Paso, El Paso, TX 79968.

The precipitation of carbides ($M_{23}C_6$) on grain boundaries in mill-processed 304 stainless steels having carbon contents of 0.01, 0.025, 0.05, and 0.07 weight percent is being investigated using transmission electron microscopy. We have compared sensitization and precipitation after aging at 625 and 775°C for more than 10^3 hours. These temperatures represent a preponderance of precipitation and healing (ex-solution) respectively, and observations of carbide density and size along similar types of grain boundaries have been correlated with these general features. Chromium depletion is also being correlated with variations in precipitate size and density. Similar boundaries were determined by comparing misorientations for identical crystallographic orientations for neighbor grains separated by the grain boundary interface. Some preliminary comparisons of precipitation on grain boundaries having different misorientations have also been made, and these observations will be described. Depletion of chromium between precipitates along selected boundaries is also being investigated using fine focussed electron beam line scans which produce line-width resolutions less than 200 Å. Work supported by NSF-RIMI Grant HRD 9105065 and EPA Cooperative Agreement CR-81 8296-01-0.

MATERIALS SCIENCES

POSTER PRESENTATION — SPRING 1993

COMPARISON OF MICROSTRUCTURES FOR COPPER AND TANTALUM SHAPED CHARGE REGIMES: H. K. SHIH, C-S. NIOU and L. E. MURR, Department of Metallurgical and Materials Engineering, The University of Texas at El Paso, El Paso, TX 79968.

The shaped charge has been used since World War II for armor penetration as well as fracturing of rock, cutting, precision separation, and demolition. A shaped charge for armor penetration consists of a conical metal liner which is surrounded by a cylindrical explosive and detonated. The detonation collapses the cone and produces a slug which contains up to 80 per cent of the cone material and an axial jet which stretches to some instability condition and particulates into necked fragments. This jetting process involves true strains in excess of 10 at strain rates between 10^4 and 10^8 s⁻¹. Initial jet-forming pressures can exceed 200 GPa. Observations of initial and residual microstructures are indicative that this example of extreme plastic deformation also appears to represent a classic example of discontinuous dynamic recrystallization. A detailed examination of microstructures which characterize the starting cone, the residual slug, and individual jet fragments (the shaped-charge regime) has been completed for copper and tantalum shaped charges and the results illustrate interesting microstructure evolution. Light microscopy and SEM provide a metallurgical overview while TEM provides details of microstructural issues on a very fine scale. Work supported by a Murchison Endowed Chair and the Phelps-Dodge Foundation through the Phelps-Dodge Scholars Program at UTEP.

ANSWER TO “WHAT IS IT”

Fractured surface of human dentin showing an area of dentin layer. “Holes” represent dentin tubules which house odontoblastic processes. Prepared by etching with 30% HCl for 30 seconds and sputter coated with gold-palladium.
(Bar = 10 micrometers)

Micrograph — Roger D. Metcalf, D.D.S., Biology Department, The University of Texas at Arlington, 76019.

APPLICATION FOR MEMBERSHIP OR CHANGE OF ADDRESS

TEXAS SOCIETY FOR ELECTRON MICROSCOPY, INC.

Date _____

Please type or print legibly. Fill out completely. The numbers in parenthesis are the maximum number of characters and spaces the computer can accommodate for that blank. Though we will mail to your home address, we prefer to have your work address. Please note that membership is for Jan. - Dec. for each year.

- Check One: ☐ I am applying for new membership in T.S.E.M.
☐ I am a member and wish to change my address.
☐ I am a STUDENT and wish to upgrade to REGULAR membership.

Name (last name first) _____ (35)

Institution _____ (35)
(Please write out completely. We'll abbreviate it.)

Department _____ (35)
(Please write out completely. We'll abbreviate it.)

Street & Number / P.O. Box _____ (35)

City _____ (20) State _____ (2) Zip _____ (10)

Work Phone (_____) _____ (13) Extension _____ (4)

Electronic Mail (_____) _____ (40)

Home Phone (_____) _____ (13) FAX No. (_____) _____ (13)

Category of Membership (circle only one): **Regular** **Corporate** **Honorary** **Library**

Student: _____ Degree Program _____ Signature of faculty sponsor _____

Broad field of interest in which you utilize Electron Microscopy (Circle only one):

Zoology	Botany	Microbiology	Cell Biology	Biochemistry
Medicine	Vet. Medicine	Chemistry	Sales	Service/Repair
Materials	Petroleum	Semiconductor	Environment	Minerals

If you are a member changing your address, please attach an old mailing label to help us identify your previous record in the computer. Applicants for membership should include a check or money order for one year's dues with application (Regular: \$15.00; Student: \$2.00; Corporate: \$75.00).

Applications for new membership, or for upgrading of membership category from STUDENT to REGULAR, will be presented to the Executive Council at their next meeting for their approval (majority vote). The applicants will then be presented by the council to the membership at the next general business meeting for their approval (majority vote). Applicants will be added to the membership rolls at that time.

Please Return To: Keith R. Fry
Secretary, TSEM
Center of Biotechnology
Baylor College of Medicine
4000 Research Forest Drive
The Woodlands, Texas 77381

800
533
2509





Our ad agency said we should promote our image.

Good idea, we said. After all, in electron microscopy, image is critical. And nobody, we told them, beats Hitachi when it comes to high-resolution images. Like the 7Å achieved with our cold field emission technology, for example.

But, we added, image isn't everything. SEMs and TEMs should be easy to use. So even non-experts always get top performance. And we explained how our computer control automates key functions and settings. How we use ergonomics to place controls conveniently, logically.

Further, we said, image is nothing without reliability. But, with our digital electronics, customers rest easy, not only about dependability, but about data accuracy, reproducibility and serviceability as well.

And what about choice, we asked. Applications vary. Budgets vary. But a good supplier can respond to any need, can offer a wide choice even in integrated imaging/microanalysis systems. Then we sent them out to count our systems—all 22 of them.

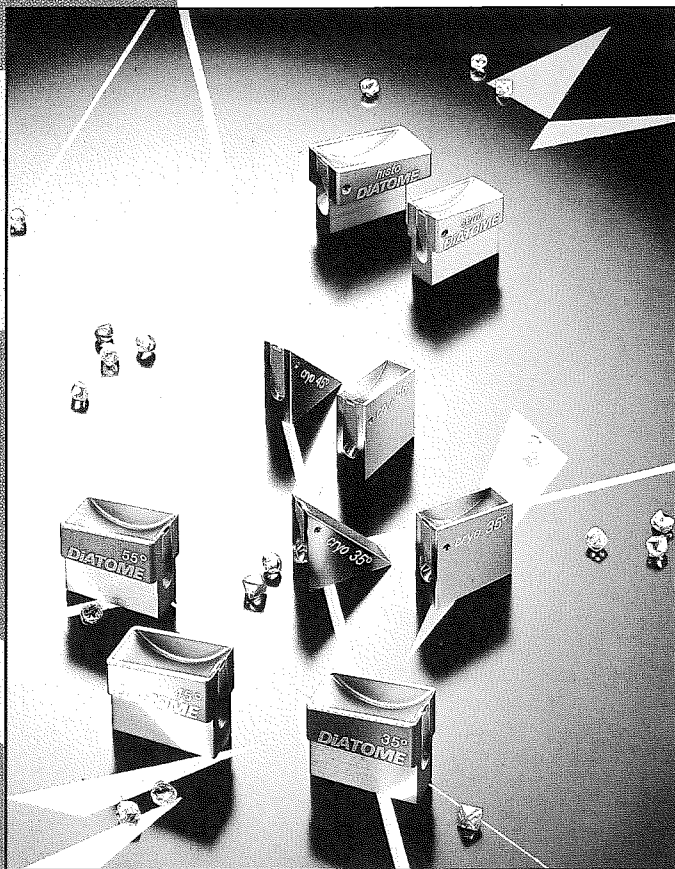
What's it all mean? Well, for one thing, the ad people learned nothing beats experience. And that when

they start talking images, we can show them a thing or two. It also means you should be getting in touch. Obviously, we're the ones to help with *your* image, too.

 **HITACHI®**
SCIENTIFIC INSTRUMENTS
Nissei Sangyo America, Ltd.

460 E. Middlefield Road
Mountain View, CA 94043
(415) 969-1100

25 West Watkins Mill Road
Gaithersburg, MD 20878
(301) 840-1650



For The Performance You Expect:

The DiATOME resharpening service.

When Diatome resharpens a Diatome Knife, we restore it to its original condition. **That is our Guarantee!** Your resharpened Diatome Knife will have the same length, the identical cutting edge and carry the same guarantee of quality as the day it first left our factory.

Only Diatome can make this claim!

No other company can successfully resharpen a Diatome Diamond Knife. We have found that when other companies try to resharpen our knives, the original parameters of our knives are either altered or totally lost (the diamond cutting edge is shorter or in some cases our diamond has been removed and replaced with a diamond of inferior quality and shorter service life). Hence, returning to you an inferior knife that does not perform as the original.

The Diatome Diamond Knife is also guaranteed for an **unlimited** number of resharpenings.

Each Diatome Diamond Knife, whether new or resharpened, is subjected to extensive testing for its ability to cut accurately without scoring or compression. Only if its performance passes our tests will we ship it to you.

This too is guaranteed!

Diatome is committed to customer satisfaction. Therefore, in the unlikely event that you experience any difficulties, or for any reason you are unhappy with the performance of your knife, please contact us immediately. You can be sure that any problem with your knife will be corrected.

We guarantee it!

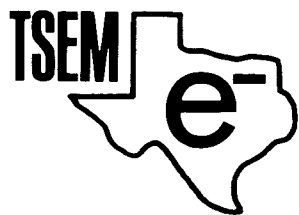
We stand by our commitment to quality and customer satisfaction.

***For Quality
For Accuracy
For Satisfaction
Forever***

DIATOME U.S.

Call or write for our complete set of literature today.
P.O. Box 125, Fort Washington, PA 19034
(215) 646-1478 • (800) 523-5874

TSEM Members



Dr. Ashok Advani, Inst. for Manufacturing & Material Management, University of Texas at El Paso, 500 W. University, El Paso, TX 79968-0520.

Howard J. Arnott, Dept. of Biology, University of Texas at Arlington, P.O. Box 19498, Arlington, TX 76019.

Rajasekaran Balasubramanian, Dept. of Mechanical Engineering, Texas A&M University, 10070 West Park #512, Houston, TX 77042.

Wayne J. Barcellona, Dept. of Biology, Texas Christian University, P.O. Box 32916, Fort Worth, TX 76129.

Steven S. Barham Ph.D., R. J. Lee Group - Houston Inc., 14760 Memorial Drive, Suite 106, Houston, TX 77079.

Roberto Barrios, Department of Pathology, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030.

Margrit Barry, Barry Scientific, Inc., P.O. Box 173, Fiskdale, MA 01518.

Paul S. Baur, Jr., 3319 Ave. R, Galveston, TX 77550.

Steven C. Bauserman, M.D., Dept. of Anatomic Pathology, Scott and White Clinic, 2401 S. 31st Street, Temple, TX 76508.

Don C. Benefiel, 103 Ligustrum, Lake Jackson, TX 77566.

Jerry Benham, Microscience Division, Bio-Rad Microscience Division, 19 Blackstone St., Cambridge, MA 02139.

Marilyn Bennett-Lilley, Semiconductor Process, Texas Instruments, Inc., P.O. Box 655012, Dallas, TX 75265.

Jan P.G. Bergmanson, College of Optometry, University of Houston, 4800 Calhoun Rd., Houston, TX 77004.

Jacques A. Berlin, Office of Public Health, New York State Dept. of Health, 584 Delaware Ave., Buffalo, NY 14202.

Jerry D. Berlin, Dean of Science and Mathematics, Southwest Missouri State University, 901 South National Ave., Springfield, MO 65804-0094.

Cole Bess, Electron Microscope Supplier Div., EMC Corp, P.O. Box 67285, Chestnut Hill, MA 02167-0004.

Graham Bird, Oxford Instruments North America, Inc., 13105 Northwest Frwy., Suite 290, Houston, TX 77040.

Virginia Blackmon, 3839 El Campo Ave., Ft. Worth, TX 76107.

Gred D. Blaisdell, 6609 Kavanaugh Place, Little Rock, AR 72207.

Yvonne Stich Blocker, College of Optometry, University of Houston, 5631 Spellman, Houston, TX 77096.

Daniel V. Blystone, Dept. of Biology, San Antonio College, 2635 Worldland, San Antonio, TX 78217.

Robert V. Blystone, Dept. of Biology, Trinity University, 715 Stadium Drive, San Antonio, TX 78284.

Doria F. Bowers, Dept. of Microbiology, University of Texas Health Science Center, 7703 Floyd Curl Drive, San Antonio, TX 78284-7758.

Louis H. Bragg, Dept. of Biology, University of Texas at Arlington, P.O. Box 19498, Arlington, TX 76019.

Bill Brinkley, Dean of Graduate Studies, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030-3498.

R. Malcolm Brown, Jr., Dept. of Botany, University of Texas at Austin, Austin, TX 78712.

Cynthia E. Brown-Duncan, Bio-Rad Microscience Division, 19 Blackstone St., Cambridge, MA 02139.

Anne-Marie Brun-Zinkernagel, Dept. of Anatomy & Cell Biology, Texas College of Osteopathic Medicine, 3500 Camp Bowie, Fort Worth, TX 76107.

Corazon D. Bucana, Dept. of Cell Biology, University of Texas - M. D. Anderson Hospital, 1515 Holcombe Blvd., HMB 173, Houston, TX 77030.

Robert Buchanan, Sales/Marketing, Topcon Tech. Inc. (Formerly ISI), 6940 Koll Center Pkwy., Pleasanton, CA 94566.

John R. Buffington, Laboratory Medicine-Microbiology, National Naval Medical Center, 8901 Wisconsin Ave., Bethesda, MD 20889.

Louis Maximilian Buja M.D., 6431 Fannin, MSB 2.137, Houston, TX 77030.

Patti Jo Burkett, P.O. Box 1258, Slidell, LA 70459.

Terry R. Burns, Dx Inc., 2203 W. Tennessee, Midland, TX 79707-3320.

James K. Butler, Dept. of Biology, University of Texas at Arlington, P.O. Box 19498, Arlington, TX 76019.

Thomas Caceci, Dept. of Veterinary Biosciences, Virginia/Maryland College of Veterinary Medicine, Virginia Polytechnic Institute, Blacksburg, VA 24061.

Edward P. Calomeni, Dept. of Pathology - EM Lab, Medical College of Ohio, P.O. Box 10008, Toledo, OH 43699-0008.

Ivan L. Cameron, Dept. of Cell. & Structural Biology, University of Texas Health Science Center, 7703 Floyd Curl Dr., San Antonio, TX 78284-7762.

Marion M. Campbell, 5609 Flack, Houston, TX 77081.

Marvin S. Cannon, Dept. of Anatomy; College of Medicine, Texas A&M University, College Station, TX 77843.

David B. Cantu-Crouch, Electron Microscopy Unit, Alcon Laboratories, 6201 South Freeway, Fort Worth, TX 76134-2099.

Tony Carpenter, Electron Optics Division, Carl Zeiss Inc., One Zeiss Dr., Thornwood, NY 10594.

Joiner Cartwright Jr., Dept. of Pathology, Room 286-A, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030.

Tapas Chanda, Dept. of Material Sciences & Engineering, The University of Michigan, 2514 Dow Bldg., Ann Arbor, MI 48109-2136.

Wah Chiu, Biochemistry, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030-3498.

Donna R. Clarkson, Dept. of Pathology, University of Texas Health Science Center, San Antonio, 7703 Floyd Curl Drive, San Antonio, TX 78284-7750.

Fred J. Clubb, Jr., D.V.M., Ph.D., Dept. of Cardiovascular Path., Texas Heart Institute, P.O. Box 20345, MC 4-265, Houston, TX 77225-0345.

Carolyn Corn, Dept. of Cell Biol. & Environm. Sci., University Health Science Center, P.O. Box 2003, Tyler, TX 75710.

Elma Cortinas, Pathology, Children's Medical Center, 1935 Motor St., Dallas, TX 75235.

Ernest F. Couch, Dept. of Biology, Texas Christian University, Fort Worth, TX 76129.

Michael Coviello, Texas Instruments - Materials Character., 13500 N. Central Expressway, P.O. Box 655936, Mail Station 147, Dallas, TX 75265.

Rick Cumby, Denton Vacuum, Inc., P.O. Box 820, Mesquite, TX 75185.

Rick Cumby, Microtek Analytical Sales, P.O. Box 820, Mesquite, TX 75185.

Linda Dailey, EMITECH U.S.A., Inc., 3845 FM 1960 West, Suite 345, Houston, TX 77068.

Marianne Dauwalder, Dept. of Botany, University of Texas, Austin, TX 78713.

Franklin D. Davidson, Physical Sciences, Pennzoil Products Co., P.O. Box 7569, The Woodlands, TX 77387.

Ronald W. Davis, Human Anatomy, Texas A&M University, College Station, TX 77843.

Glenn L. Decker, Dept. Biochem & Molecular Bio., Box 117, University of Texas - M. D. Anderson Cancer Center, 1515 Holcombe Blvd., Houston, TX 77030.

Anthony Del Campo, Life Cell Corporation, 3606-A Research Forest Drive, Woodlands, TX 77381.

Ms. Andrew Devine, Marketing, FEI Co., 19500 N. W. Gibbs Dr., Suite 100, Beaverton, OR 97006-6907.

Joe B. Dixon, Dept. of Soil & Crop Sciences, Texas A&M University, College Station, TX 77843-2474.

Sam Dominey, P.O. Box 423, Dodge, TX 77334.

C. Delfina Dominguez, Department of Biological Sciences, University of Texas at El Paso, 3023 Piedmont, El Paso, TX 79902.

Lisa Donaghe, EM Center, Texas A&M University, MS #2257, College Station, TX 77843.

Jonnell Drab, Electron Microscopy Unit, Alcon Laboratories, 6201 South Freeway R5-12, Fort Worth, TX 76134-2099.

Bob Droleskey, Vet. Toxicol. & Entomol. Res. Lab, United States Dept. of Agriculture, Rt. 5 Box 810, College Station, TX 77840.

Douglas Carl Dufner, EM Center, Texas A&M University, College Station, TX 77843.

E. Steven Duke, Outpatient Clinic/Clinical Research, University of Texas Health Science Center, 7703 Floyd Curl Drive, San Antonio, TX 78284-7914.

Kenneth Dunner, Jr., Dept. of Cell Biology, University of Texas - M.D. Anderson Cancer Center, 10950 Westbrae Parkway, Apt. #6210, Houston, TX 77031.

Dena M. Edwards-Stark, Dept. of Pathology, University of Texas Health Science Center - San Antonio, 7703 Floyd Curl Dr., San Antonio, TX 78284-7750.

Joanne Tontz Ellzey, Dept. of Biological Sciences, University of Texas at El Paso, Ultrastructure Laboratory, El Paso, TX 79968-0519.

Rolando Estrada-Gordillo, Department of Pathology, St. Elizabeth Hospital, 2830 Calder Ave., Beaumont, TX 77702.

- Margaret L. Farley**, Dept. of Pathology; Cytopath. Lab, University of Texas Health Science Center, P.O. Box 2003, Tyler, TX 75710.
- John Fitzpatrick**, Emitech, 3845 FM 1960 West, Suite 345, Houston, TX 77068.
- Jesse Flores**, Garrett Laboratories, 3718 Galloway Lane, Carrollton, TX 75007.
- Keith R. Fry**, Center for Biotechnology, Baylor College of Medicine, 4000 Research Forest Dr., The Woodlands, TX 77381.
- Charles A. Garber**, Div. of Structure Probe Inc., SPI Supplies, P.O. Box 656, 569 E. Gay St., West Chester, PA 19381-0656.
- David Garrett**, 713 Biggs Terrace, Arlington, TX 76010.
- Susan L. Garrett**, Department of Anthropology, Southern Methodist University, Dallas, TX 75275.
- William Geoghegan**, Dept. of Dermatology, University of Texas Medical School, 6431 Fannin, MSMB 1.204, Houston, TX 77030.
- Catherine J. Giddings**, Dept. of Cell Biology, University of Texas Southwest Medical Center, 5323 Harry Hines, Dallas, TX 75235-9039.
- Patricia Glazebrook**, Center for Biotechnology, Baylor College of Medicine, 4000 Research Forest Dr., Woodlands, TX 77386.
- David Glennon**, AMRAY Inc., 8178 W. Peakview Drive, Littleton, CO 80123.
- Jan Goddard-Finegold**, Texas Children's Hospital, Clinical Care Center, MC3-3311, 6621 Fannin St., Houston, TX 77030.
- Margaret Ann Goldstein**, Dept. of Medicine, Cardiovasc. Sci., Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030.
- William J. Gonyea, Ph.D.**, Dept. of Cell Biology & Neuroscience, University of Texas Health Science Center, 5323 Harry Hines Blvd., Dallas, TX 75235-9039.
- M. Lynn Gray**, Dept. of Cell Bio. & Environ. Sci., University of Texas Health Science Center, Box 2003, Tyler, TX 75710.
- Sy Griffey**, Center for Biotechnology, Baylor College of Medicine, 4000 Research Forest Drive, The Woodlands, TX 77381.
- Lawrence R. Griffing**, Biology, Texas A&M University, College Station, TX 77843.
- Donald L. Grimes**, 2227 Branch St., Middleton, WI 53562.
- John V. Grimes**, Dept. of Biology, Midwestern State University, Wichita Falls, TX 76308-2099.
- John R. Guyton**, Dept. of Medicine, Mail Station A-601, Baylor College of Medicine, 6565 Fannin St., Houston, TX 77030.
- Candace H. Haigler Ph.D.**, Department of Biological Sciences, Texas Tech University, Box 4149, Lubbock, TX 79409-3131.
- David B. Halpern**, P.O. Box 4040, Rydel, PA 19046.
- Hal K. Hawkins**, Dept. of Pathology, Texas Children's Hospital, Baylor College of Medicine, P.O. Box 20269, Houston, TX 77030-0269.
- Don A. Hay**, Dept. of Biology, Stephen F. Austin State University, P.O. Box 13003, Nacogdoches, TX 75962-3003.
- Tetsuya Hayashi**, World Health Organization - Cardiovascular Center, University Texas Medical Branch, Room 2.110, Brackenridge H-80, Galveston, TX 77550.
- Elizabeth S. Hayes**, University of Texas, Box 227, Galveston, TX 77550-2783.
- Julian P. Heath**, Department Pediatrics - CNRC, Baylor College of Medicine, 1100 Bates, Houston, TX 77030.
- Karin Westlund High**, Dept. of Anat & Neurosciences, University of Texas Medical Branch, 200 University, Galveston, TX 77550.
- Terrell R. Hoage**, Div. of Life Science-Geol. Geog., Sam Houston State University, P.O. Box 2116, Huntsville, TX 77341.
- Meg Hoobler**, Microscopy Dept., Polysciences Inc., 400 Valley Rd., Warrington, PA 18976.
- John C. Irmen**, Irmen Technical Sales, Rt. 626, Viewtown, VA 22746.
- Sally Jackson**, Dept. of Biology, Baylor University, P.O. Box 97388, Waco, TX 76798-7388.
- Sheila R. Jeffcoat**, Dept. of Biology, Southwest Texas State University, 1624 Aquarena Springs Dr., G-149, San Marcos, TX 78666-4616.
- Edwin H. Johnson**, Lab Service, Olin E. Teague Veterans' Center - 113, 1901 South First St., Temple, TX 76504.
- Gretchen D. Jones**, Dept. of Botany, Texas A&M University, College Station, TX 77843-3258.
- Stanley D. Jones**, Dept. of Rangeland Management & Ecology, Texas A&M University, College Station, TX 77843.
- Mary Faggard Kanz**, Dept. of Pathology, University of Texas Medical Branch, 230 Keiller Bldg., Galveston, TX 77550.
- A. Stephen Kester**, Dept. of Biological Science, North Texas State University, Denton, TX 76203.
- R. Thomas King**, Dept. of Surgical Pathology, Scott & White Clinic, 2401 South 31st St., Temple, TX 76508.
- Glenn Kinnear**, Oxford Instruments North America, Inc., 3-A Alfred Circle, Bedford, MA 01730.
- Eric W. Kirkland**, Bal-Tec Products, 166 Sunset Terrace, Laguna Beach, CA 92651.
- C. Ward Kischer**, Dept. of Anatomy, University of Arizona College of Medicine, Health Science Center, Tucson, AZ 85724.
- Marcella M. Klima, M.D.**, 2039 MacArthur, Houston, TX 77030.
- Joseph R. Koke**, Department of Biology, Southwest Texas State University, San Marcos, TX 78666.
- Stephen J. Kolodziej**, Dept. of Pathology, University of Texas Health Science Center, P.O. Box 20708, Houston, TX 77225.
- Sheri Kurland/Larry Kolodziejski**, Gatan Inc., 6678 Owens Dr., Pleasanton, CA 94588-3334.
- Laszlo G. Komuves**, Children's Nutrition Res. Center, Baylor College of Medicine, 1100 Bates St., Houston, TX 77030-2600.
- Cynthia A. Koutz**, Department of Ophthalmology, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030.
- Dimitrij Lang**, Retired, 802 St. Lukes Drive, Richardson, TX 75080.
- Paulette Langlinais**, Dept. of Pathology, U.S. Army Inst. of Surgical Res., Bldg. 2653, Ft. Sam Houston, TX 78234-5012.
- Laurie Chudej Leer**, Dept. of Pathology, Texas Children's Hospital, P.O. Box 20269, Houston, TX 77225.
- Luther E. Lindner**, Dept. of Pathology & Lab Med., Texas A&M University, College of Medicine, College Station, TX 77843.
- Richard Lois**, JEOL (U.S.A) Inc., 3714 Swift Creek Dr., Kingwood, TX 77339.
- James C. Long**, EM Center, Texas A&M University, College Station, TX 77843-2257.
- Jo L. Long**, Philips Electronic Instruments Inc., 6575 West Loop South, Suite 270, Bellaire, TX 77401.
- Ray Loxterman**, Rayco Photo Equipment Services, Inc., 4800 W. 34th St., Suite C-53, Houston, TX 77092.
- Newell McArthur**, Dept. of Veterinary Anatomy, Texas A&M University, College Station, TX 77843.
- Mitchell D. McCartney**, EM Unit, RO-11, Alcon Laboratories, Inc., 6201 South Freeway, Ft. Worth, TX 76134-2099.
- Linda M. McClellan**, 2517 Holt Rd., Arlington, TX 76006.
- William B. McCombs, III**, Dept. of Microbiology & Immunology, Scott & White Clinic, Temple, TX 76501.
- Cameron Elaine McCoy**, Virology Section, Scott & White Clinic, 2401 S. 31st Street, Temple, TX 76508.
- Kathlyn Steer McElveen**, 3005 Longbow, Garland, TX 75042.
- Linda M. McManus**, Dept. of Pathology, University of Texas Health Science Center, 7703 Floyd Curl Dr., San Antonio, TX 78284-7750.
- Bruce MacKay**, Dept. of Anatomic Pathology, University of Texas - M.D. Anderson Hospital, 6723 Bertner Ave., Rm. G-704, Houston, TX 77030.
- Camelia Maier**, Dept. of Biological Sciences, University of North Texas, P.O. Box 5218, Denton, TX 76203.
- Peter Brian Marcus M.D.**, Dept. of Pathology, Methodist Medical Center, 1441 N. Beckley, Dallas, TX 75203.
- Joe A. Mascorro**, Dept. of Anatomy, Tulane Medical School, 1430 Tulane Ave., New Orleans, LA 70112.
- Dixie Matlock**, Dept. of Metal. and Mat. Eng., University of Texas at El Paso, 500 University, El Paso, TX 79912.
- John M. Mendenhall**, Dept. of Botany, University of Texas at Austin, BIO 308, Austin, TX 78713.
- Phebe (Clark) Mertes**, Texas A&M University, 332A Reynolds Medical Bldg., College Station, TX 77843.
- Bernard E. Mesa/Cathy Zimmerman**, Micro Engineering Inc., Rt. 2, Box 474, Huntsville, TX 77340.
- Rob Meyer/Mathew Batchelor**, Meyer Instruments, Inc., 1304 Langham Creek Drive, Suite 235, Houston, TX 77084-5042.
- Abdul Jabbar Mia**, Prof. & Chair, Division of Science & Mathematics, Jarvis Christian College, Hwy. 80, Hawkins, TX 75765-9989.
- William I. Miller, III**, Bal-Tec Products Inc., 984 Southford Road, Middlebury, CT 06762.
- Richard E. Miller**, Electron Microscope Center, Texas A&M University, College Station, TX 77843-2257.
- Margaret Mary (Peggy) Miller**, Dept. of Pathology, University of Texas Health Science Center, 7703 Floyd Curl Drive, San Antonio, TX 78284-7750.
- Charles W. Mims**, Dept. of Plant Pathology, University of Georgia, Athens, GA 30602.
- Janet I. Minshew**, Leica, Inc., 3905 Inverness Ln., Plano, TX 75075.
- Hilton H. Mollenhauer**, 1208 N. Ridgefield Circle, College Station, TX 77840-4339.
- Ellen G. Moore**, Dept. of Veterinary Toxicology, USDA-ARS-FAPRL, Rt. 5 Box 810, College Station, TX 77845.
- Randy Moore**, Dept. of Biological Sciences, Wright State University, Dayton, OH 45435.
- Donald Foster Mullica**, Dept. of Chemistry, Baylor University, P.O. Box 7431, Waco, TX 76798.
- Lawrence E. Murr**, Metallurgical & Materials Eng., University of Texas at El Paso, El Paso, TX 79968.
- Hideo Naito**, Hitachi Scientific Instruments Division, 460 E. Middlefield Rd., Mountain View, CA 94043.
- Pamela J. Neill**, The Center for Biotechnology, Baylor College of Medicine, 4000 Research Forest Dr., The Woodlands, TX 77381.
- David M. Nidosik**, Dept. Forest Sc/Coll Agriculture, Texas A&M University, Box 2135, College Station, TX 77843-2135.
- Zhu Hua Ning**, Division of General Study, Jarvis Christian College, P.O. Drawer G, Hawkins, TX 75765.
- Kimberly L. Novak**, 2137 Park Springs Circle #1082, Arlington, TX 76013.

- Lawrence X. Oakford, Ph.D.**, Dept. of Anatomy & Cell Biology, Texas College of Osteopathic Med, 3500 Camp Bowie, Fort Worth, TX 76107-2690.
- Udayan K. Parikh**, Anatomical Sciences, UTHSC-Houston Dental Branch, 6516 John Freeman Ave., Suite 4.099, Houston, TX 77030.
- Katherine Patterson**, Lon Morris College, P.O. Box 1562, Jacksonville, TX 75766.
- Ted Pella**, Ted Pella, Inc., P.O. Box 2318, Redding, CA 96099.
- Michael W. Pendleton**, Dept. of Anthropology, Texas A&M University, College Station, TX 77843.
- Phillip Perlman**, Electron Microscopy Laboratory, Don & Sybil Harrington Cancer Center, 1500 Wallace, Amarillo, TX 79106.
- Dan Petr**, Department of Biology, Southwestern Adventist College, Hillcrest and Mockingbird, Keene, TX 76059.
- Charles W. Philpott**, Dept. of Biology, Rice University, Houston, TX 77001.
- Alfred Pick**, ElectroScan Corp., 6345 Douglas Street #161A, Plano, TX 75093.
- Gerald J. Pinero**, Dept. of Anatomical Sciences, University of Texas Health Science Center - Dental Branch, P.O. Box 20068, Houston, TX 77225.
- Russell F. Pinozzotto**, Center for Materials Characterization, University of North Texas, P.O. Box 5308, Denton, TX 76203.
- Pamela J. Potts**, Dept. of Cardiovascular Path., Texas Heart Institute, P.O. Box 20345, MC4-265, Houston, TX 77225-0345.
- Cyrus Pouraghabagher**, Center for Materials Characterization, University of North Texas, UNT Box 6955, Denton, TX 76203.
- Sheila Powell**, MOS 8, Motorola, Inc., 912 Silver Quail, Austin, TX 78753.
- Kellous A. Price**, Dept. of Biology, Southwest Texas State University, San Marcos, TX 78666.
- Nan Webb Pryor**, USDA-FAPRL, Rt. 5, Box 810, College Station, TX 77840-9594.
- Lew Rabenberg**, Dept. of Mech. Eng., University of Texas at Austin, Materials Sci. & Eng., Austin, TX 78712-1063.
- Elsa E. Ramos**, Dept. Pathology, M.D. Anderson Hospital, 6723 Bertner, Rm G-704, Houston, TX 77030.
- Jane Ramberg**, Dept. of Internal Medicine, University of Texas Health Science Center, 5323 Harry Hines Blvd., Dallas, TX 75235-8884.
- Ruben Ramirez-Mitchell**, The Cell Research Institute, University of Texas, P.O. Box 7640, Austin, TX 78713-7640.
- Richard Rebert/Stacie Kirsch**, Sales Department, Electron Microscopy Sciences, 321 Morris Rd., P.O. Box 251, Fort Washington, PA 19034.
- Young G. Rho**, Department of Physics, University of North Texas, 2413 W. Prairie, Apt. 20, Denton, TX 76201.
- Hobie Richards**, EDAX International Inc., 2316 Stoney Brook Lane, Flower Mound, TX 75028.
- Mildred Richards**, EM Center, Texas A&M University, BSBW 129, College Station, TX 77843-2257.
- Ezquiel R. Rivera**, Dept. of Biol. Sci., University of Massachusetts - Lowell, 1 University Avenue, Lowell, MA 01854.
- Daniel K. Roberts**, Dept. of Obstetrics and Gynecology, University of Kansas School of Medicine, Wesley Medical Center, 550 N. Hillside, Wichita, KS 67214.
- David A. Roberts**, A. Lischen Kerr/Len Strickland, RMC Inc., 4400 S. Santa Rita Ave., Tucson, AZ 85714.
- David A. Roberts**, Basic Research, Sid Richardson Carbon and Gasoline, 4825 N. Freeway, Fort Worth, TX 76137.
- Susan E. Robbins**, Dept. of Pathology, Rm. 212-B, Baylor College of Medicine, 1200 Moursund, Houston, TX 77030.
- Melody K. Robinson**, The D & S Harrington Cancer Center, 1500 Wallace Blvd., Amarillo, TX 79106.
- Lynn Rodgers**, Biology, Texas A&M University, 1206A Holik, College Station, TX 77840.
- Wendy Belinda Rosser**, Department of Biology, Baylor University, P.O. Box 97388, Waco, TX 76798-7388.
- M. Lea Rudee**, Dean of Engineering, University of California at San Diego, La Jolla, CA 92037.
- Ann E. Rushing**, Biology, Baylor University, P.O. Box 97388, Waco, TX 76798-7388.
- H. Wayne Sampson**, Dept. of Human Anatomy, Texas A&M University Medical School, College Station, TX 77843.
- Barbara Schneider**, Department of Pathology, University of Texas Health Science Center at San Antonio, 7703 Floyd Curl Drive, San Antonio, TX 78284-7750.
- John P. Schroeter**, Dept. Ecology & Evol. Biology, Rice University, Houston, TX 77251.
- Betty Lou Schumaker**, 1321 E. 279th St., Apt. 204C, Euclid, OH 44132.
- Mary Schunder Ph.D.**, Dept. of Anatomy, Texas College of Osteopathic Medicine, 3500 Camp Bowie Blvd., Ft. Worth, TX 76107-2690.
- Linda Schutze**, Research and Development, LifeCell Corporation, 3606 Research Forest Drive, The Woodlands, TX 77381.
- Randy Scott**, Electron Microscope Center, Texas A&M University, College Station, TX 77843-2259.
- Lydia Shanks**, Dept. Pathology/EM Lab, M.D. Anderson Hospital, 1515 Holcombe, Houston, TX 77030.
- D. Janine Sherrier**, Dept. of Biology, Texas A&M University, College Station, TX 77843.
- Leo D. Simone**, State University of New York, College of Potsdam, Potsdam, NY 13676.
- Perry K. Simons**, Dept. of Pathology, St. John Medical Center, 1923 South Utica Ave., Tulsa, OK 74104.
- Helge Sittertz-Bhatkar**, EM Center, Texas A&M University, 2913 Rayadu, College Station, TX 77845.
- Harold Samuel Skjonsby Ph.D.**, Dept. of Anatomical Sciences, University of Texas Health Science Center at Houston - Dental Branch, P.O. Box 20068, Houston, TX 77225.
- Steven E. Slap**, Energy Beam Sciences, Inc., P.O. Box 468, 11 Bowles Rd., Agawam, MA 01001.
- G. Con Smith**, Dept. of Virology & Immunology, Southwest Foundation for Biomedical Research, P.O. Box 28147, San Antonio, TX 78284.
- Don W. Smith**, Department of Biological Sciences, University of North Texas, UNT Box 5218, Denton, TX 76203-5218.
- Nancy K. R. Smith**, Dept. of Cellular & Structural Biol., University of Texas Health Science Center, 7703 Floyd Curl Drive, San Antonio, TX 78284-7762.
- Robert Spears**, Dept. of Anatomy, Baylor College of Dentistry, 3302 Gaston Ave., Dallas, TX 75246.
- Mannie C. Steglich**, Dept. of Pathology, University of Texas - M.D. Anderson Hospital, 1515 Holcombe, Mail Box 85, Houston, TX 77030.
- James R. Stewart**, Dept. of Biology, University of Texas at Tyler, 3900 University Blvd., Tyler, TX 75701-6699.
- Mary K. Sullivan**, SCANNING/FACMS Inc., P.O. Box 832, Mahwah, NJ 07430.
- Joseph W. Tabeling**, Delaware Diamond Knives, 3825 Lancaster Pike, Wilmington, DE 19805.
- Alan Taylor**, Dept. of Microanatomy, Baylor College of Dentistry, 3302 Gaston Ave., Dallas, TX 75246.
- Josephine Taylor**, Dept. of Biology, Texas A&M University, College Station, TX 77843.
- Carl F. Tessmer**, Route 5, Box 5291, Belton, TX 76513.
- E. L. Thurston**, Electron Microscopy Center, Texas A&M University, 4905 Afton Oaks, Box 288, Route 3, Bryan, TX 77801.
- Douglas Tieden**, Department of Biology, Southern Methodist University, 200 Fondren Science, Dallas, TX 75275.
- Don A. Tiffin**, Materials Engineering, Advanced Micro Devices, 5517 Rice Drive, Box 97902, The Colony, TX 75056.
- Dan Timmermann Jr.**, Dept. of Biological Sciences, Arkansas State University, P.O. Box 787, State University, AR 72467.
- Tracy Toliver**, Southwest Texas State University, 900 Peques #1005, San Marcos, TX 78666.
- Stanley E. Trauth**, Dept. of Biological Sciences, Arkansas State University, P.O. Box 599, State University, AR 72467-0599.
- Robert A. Turner**, 3914 Erie, Temple, TX 76501.
- Kathryn A. VandenBosch**, Biology, Texas A&M University, College Station, TX 77843.
- Regina Verani M. D.**, Dept. of Pathology, University of Texas Medical School, P.O. Box 20708, Houston, TX 77225.
- Allen Waldo**, Dept. of Biology, Texas Woman's University, P.O. Box 23971, Denton, TX 76204-1971.
- Nola Busby Walker**, Dept. of Obstetrics-Gynecology [EM], Wesley Medical Center, 550 N. Hillside, Wichita, KS 67214.
- Ron Warren**, MOS 8, Motorola, Inc., 752-A Simonetti, Austin, TX 78748.
- Melvin Watson**, 2151 Baringer, Louisville, KY 40204.
- William Wehling**, NORAN, 112 Shady Oak Drive, Georgetown, TX 78628.
- Julie W. Wen**, Dept. of Pathology Route F01, University of Texas Medical Branch at Galveston, 234 Keiller Bldg., Galveston, TX 77550.
- Richard Allen White**, Materials & Interface Char., SEMATECH, 7808 Peaceful Hill, Austin, TX 78748.
- Paula S. Williamson**, Biology, Southwest Texas State, San Marcos, TX 78666.
- John N. Williard**, Betz Laboratories, 9669 Grogans Mill Road, Woodlands, TX 77380.
- Ted Willmarth**, DiaTECH, Inc., 1209 Dogwood Drive, Kingston, TN 37763.
- Zygfried R. Wolanski**, PE20116, 3520 Lawndale Avenue, Ft. Worth, TX 76133-3017.
- Joe G. Wood, Ph.D.**, Dept. of Anatomical Sciences, University of Oklahoma Health Science Center, Oklahoma City, OK 73104.
- Robert D. Yates**, Dept. of Anatomy, Tulane University Medical Center, 1430 Tulane Ave., New Orleans, LA 70112.
- Yingren (Jessica) Zhu**, EM Lab - Pathology, Texas Children's Hospital, P.O. Box 20269, Houston, TX 77225-0269.

WANTED

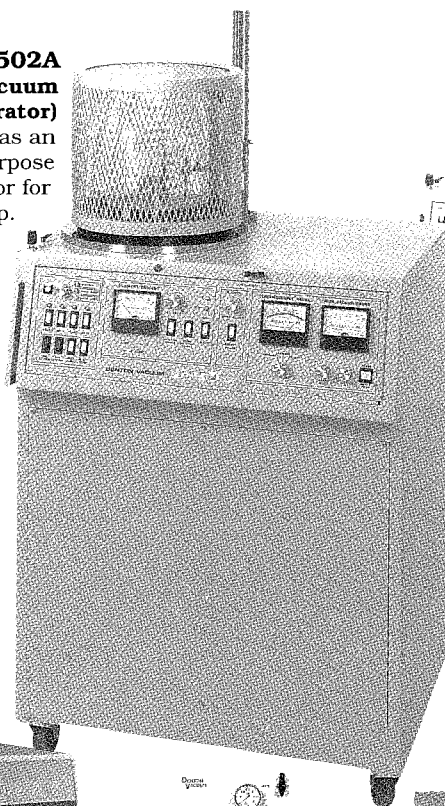
For All Sample Prep Jobs

THE DENTON GANG

DV-502A (AKA High-vacuum Evaporator)

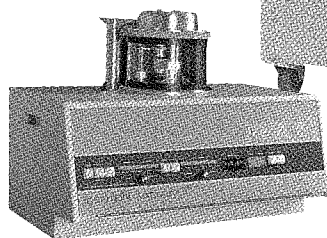
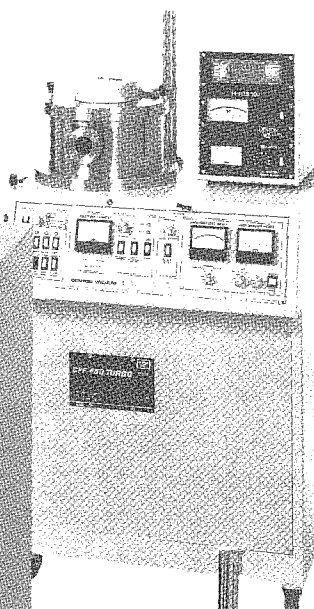
Wanted as an economical, general purpose high-vacuum evaporator for SEM/TEM sample prep.

Standard processes: carbon and thermal evaporation. Known to have extensive list of options for additional applications. Turbomolecular and cryogenic pumped versions are also available as enhancements to basic system.



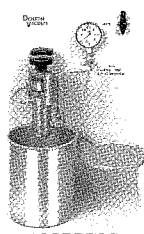
HI-RES 100 (AKA High-resolution Chromium Deposition System)

Continues to meet sample prep needs of today's high resolution field emission SEMs. With superior film quality and low sample contamination. Produces controlled, ultrathin (10Å) high purity films of Cr, Ta, Pt, etc. High-vacuum capability of 10^{-7} torr with excellent water vapor pumping speed. Uses a 150 liter per second turbomolecular pump and integral LN_2 trap.



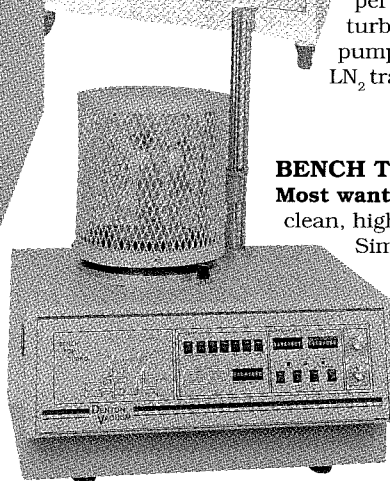
DESK II (AKA Cold Sputter Unit with Etch Mode)

Known leader of the industry. Pumpdown-coating-venting in 3 minutes. The coolest sputter coater on the market — uses a magnetron sputter head and a patented anode grid to minimize substrate heating.



CRITICAL POINT DRYER

Wanted as an Accessory: Simple, Economical and efficient. **MO:** Uses Freon or Liquid CO_2



BENCH TOP TURBO

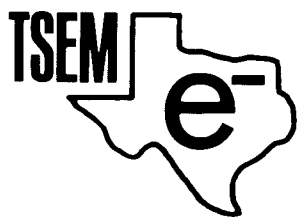
Most wanted for delivering a clean, high (10^{-9}) vacuum. Simple, one-button pump down operation without the need for air or water utilities. Large 10" x 12" bell jar facilitates the installation of multiple accessories.

Processes include carbon or metal evaporation... and sputtering of gold, gold palladium, titanium, etc. with optional accessories.

**DENTON
VACUUM
INC**

2 Pin Oak Lane, Cherry Hill, NJ 08003-4072
Phone: (609) 424-1012 Fax: (609) 424-0395

CORPORATE MEMBERS



AMRay, Inc., David Glennon. 8178 W. Peaview Dr., Littleton, CO., 80123. (214) 933-5514; (303) 933-7122.

Bal-Tec Products, Eric W. Kirkland. 166 Sunset Terrace, Laguna Beach, CA 92651. (203) 598-3660. FAX (714) 494-3551.

Barry Scientific, Inc., Margrit Barry. P.O. Box 173, Fiskdale, MA 01518. (508) 347-9855; (800) 348-9855.

Bio-Rad Microscience Division, Cynthia E. Brown-Duncan. 19 Blackstone St., Cambridge, MA 02139. (617) 864-5820; (800) 444-1422.

Delaware Diamond Knives, Joseph W. Tabeling. 3825 Lancaster Pike, Wilmington, DE 19805. (302) 999-7476; (800) 222-5143. FAX (302) 999-8320.

DiaTECH, Inc., Ted Willmarth. 1209 Dogwood Dr., Kingston, TN 37763. (615) 376-6358. FAX (615) 938-2721.

EDAX International, Robert (Hobie) Richards. 2316 Stoney Brook Lane, Flower Mound, TX 75028. (214) 539-6633; FAX (214) 219-0891.

ElectroScan Corp., Alfred Pick. 6345 Douglas St., Suite 161A, Plano, TX 75093. (214) 250-6663.

Electron Microscopy Sciences/Diatome U.S., Stacie Kirsch/Richard Rebert. 321 Morris Road, P.O. Box 251, Fort Washington, PA 19034. (215) 646-1566; (800) 523-5874. FAX (215) 646-8931.

EM Corp., Cole Bess, Electron Microscope Supplies Division. P.O. Box 67285, Chestnut Hill, MA 02167-0004. (617) 969-5583.

EMITECH U.S.A., Inc., Linda Dailey. 3845 FM-1960 West, Suite 345, Houston, TX 77068. (713) 893-2067.

Energy Beam Sciences, Inc., Steven E. Slap. P.O. Box 468, 11 Bowles Rd., Agawam, MA 01001. (413) 786-9322. FAX (413) 789-2786.

FEI Co., Andree Devine, Marketing. 19500 N.W. Gibbs Dr., Suite 100, Beaverton, OR 97006-6907. (503) 690-1520. FAX (503) 690-1509.

Gatan Inc., Sheri Kurland/Larry Kolodziejski. 6678 Owens Dr., Pleasanton, CA 94588-3334. (510) 463-0200.

Hitachi Scientific Inst., John Fitzpatrick. 3845 FM 1960 W., Suite 345, Houston, TX 77068. (713) 893-2067; (713) 893-0494.

Hitachi Scientific Instruments Division, Nissei Sangyo America, Ltd., Hideo Naito. 460 E. Middlefield Rd., Mountain View, CA 94043. (415) 961-0461; (800) 227-8877.

Irmen Technical Sales, John C. Irmen. Rt. 626, Box 267A1, Viewtown, VA 22746. (703) 937-4290. FAX (703) 937-4290.

JEOL (U.S.A.), Inc., Richard Lois. 3503-A Cedar Knolls, Kingwood, TX 77339. (713) 358-2121; (713) 358-4417.

Leica, Inc., Janet Minschew. 3905 Inverness Lane, Plano, TX 75075. (214) 985-0981; (214) 343-3769. FAX (708) 405-8139.

Meyer Instruments, Inc., Rob Meyer/Matthew Batchelor. 1304 Langham Creek Drive, Suite 235, Houston, TX 77084-5042. (713) 579-0342. FAX (713) 579-1551.

Micro Engineering Inc., Bernard E. Mesa/Cathy Zimmerman. Rt. 2, Box 474, Huntsville, TX 77340. (409) 291-6891; 1-800-533-2509. FAX (409) 294-9861.

Molecular Dynamics (Sarastro), William Miller. 21 Berkshire Blvd., Bethel, CT 06801, (203) 730-0941; (800) 635-2873.

Noran Instruments, Inc., William Wehling. 112 Shady Oak Drive, Georgetown, TX 78628. FAX (608) 836-7224.

Oxford Instruments North America, Inc., Glenn Kinnear. 3A Alfred Circle, Bedford, MA 01730. (617) 275-4350.

Oxford Instruments, Graham R. Bird, Regional Sales Manager. 13105 Northwest Frwy., Suite 290, Houston, TX 77040. (713) 462-0200. FAX (713) 462-0233.

Ted Pella, Inc., Ted Pella, President. P.O. Box 2318, Redding, CA 96099. (916) 243-2200; (800) 237-3526.

Philips Electronic Instruments, Inc., Jo Long. 6575 West Loop South, Suite 270, Bellaire, TX 77401. (713) 668-5004. FAX (713) 668-5567.

Polysciences, Inc., Meg Hoobler, Microscopy Dept. 400 Valley Rd., Warrington, PA 18976. (215) 343-6484.

Rayco Photo Equipment Services, Inc., Ray Loxterman. 4800 W. 34th St., Suite C-53, Houston, TX 77092.

RMC Inc., David Roberts/A. Lischen. Kerr/Len Strickland. 4400 S. Santa Rita Ave., Tucson, AZ 85714. (602) 889-7900. FAX (602) 741-2200.

Scanning/FACMS, Inc., Mary K. Sullivan. The JBI Building, Box 832, Mahwah, NJ 07430. (201) 818-1010. FAX (201) 818-0086.

SPI Supplies, Division of Structure Probe, Inc., Charles A. Garber. P.O. Box 656, 569 E. Gay St., West Chester, PA 19381-0656. (800) 242-4774. FAX (215) 436-5755.

Topcon Technologies, Inc., Robert Buchanan, Sales/Marketing. 6940 Koll Center Parkway, Pleasanton, CA 94566. (510) 462-2212. FAX (510) 462-2234.

Topometrix, Rick Cumby. P.O. Box 820, Mesquite, TX 75185. (214) 289-0011. FAX (214) 289-0011.

Carl Zeiss, Inc., Electron Optics Division, Tony L. Carpenter, District Sales Manager. 6809 Ragan Drive, The Colony, TX 75056. (214) 625-8525. FAX (914) 681-7443.

**MAKE PLANS NOW TO ATTEND THE
FALL, 1993 MEETING OF THE TSEM
October ★ Galveston, Texas**

DENKA LaB6

DENKA now offers an even wider selection of LaB6 cathode styles to suit your specific electron beam applications

MODEL 7
Hyper-Beam

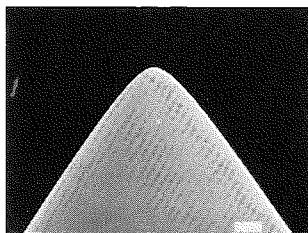
Leading electron microscope manufacturers, such as AMRAY, Cambridge, ElectroScan, ISI, JEOL and Philips have selected DENKA LaB6 as their perfect high-resolution electron beam sources. Now, in addition to the DENKA Model 3 Cathode, which is recognized as the standard in the industry for brightness and long life, DENKA announces the introduction of the new Model 7 Cathode *Hyper-Beam*, which offers unsurpassed stability without sacrificing brightness.

DENKA LaB6 Cathodes are available with any of three standard tip configurations, offering the widest range of choices to the microscopist.

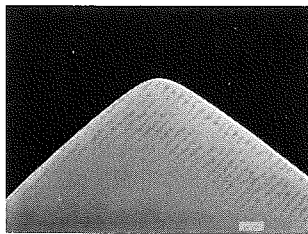
SHARP TIP: The sharp tip ($<100> 60^\circ$ cone angle, 10μ tip radius) is recommended for applications requiring the highest brightness. It is particularly effective for X-ray analysis of microscopic areas below 15A , and for many TEM applications.
ROUND TIP: The round tip ($<100> 90^\circ$ cone angle, 15μ tip radius) features a balanced combination of high brightness and long life, and is the appropriate choice for most SEM applications.

FLAT TIP: The flat tip ($<100> 90^\circ$ cone angle, 20μ tip diameter) offers the longest service life, the maximum stability and is the simplest to use. It is well-suited for many industrial applications where highest brightness is not a critical factor.

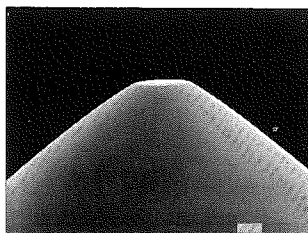
In addition to these standard tip designs, DENKA can manufacture a wide variety of custom tip shapes for your individual needs.



SHARP TIP



ROUND TIP



FLAT TIP

DENKA

DENKI KAGAKU KOGYO KABUSHIKI KAISHA
4-1, Yuraku-cho 1-chome, Chiyoda-ku,
Tokyo 100, Japan
Telephone: Tokyo 3507-5268

For inquiries, please contact:

U.S.A. & CANADA

Energy Beam Sciences
P.O. Box 468, 11 Bowles Road, Agawam, MA 01001
Tel: Toll Free 800-992-9037 Fax: (413) 789-2786
(Importer)
Mitsui Plastics Inc.
1-11, Martine Ave. White Plains, NY 10606
Tel: (914) 287-6831 Fax: (914) 287-6850

EUROPE

Dusseldorf: MITSUI & CO. DEUTSCHLAND GmbH
(DUSCP Sect.)

4000 Dusseldorf 1, Königsallee 92a, F.R. GERMANY
Tel: (211) 8796-246 Fax: (211) 8798-268

München: MITSUI & CO. DEUTSCHLAND GmbH
(MUNZZ Sect.)

8000 München 40, Leopoldstrasse 19, F.R. GERMANY,
Tel: (89) 397021 Fax: (89) 336820

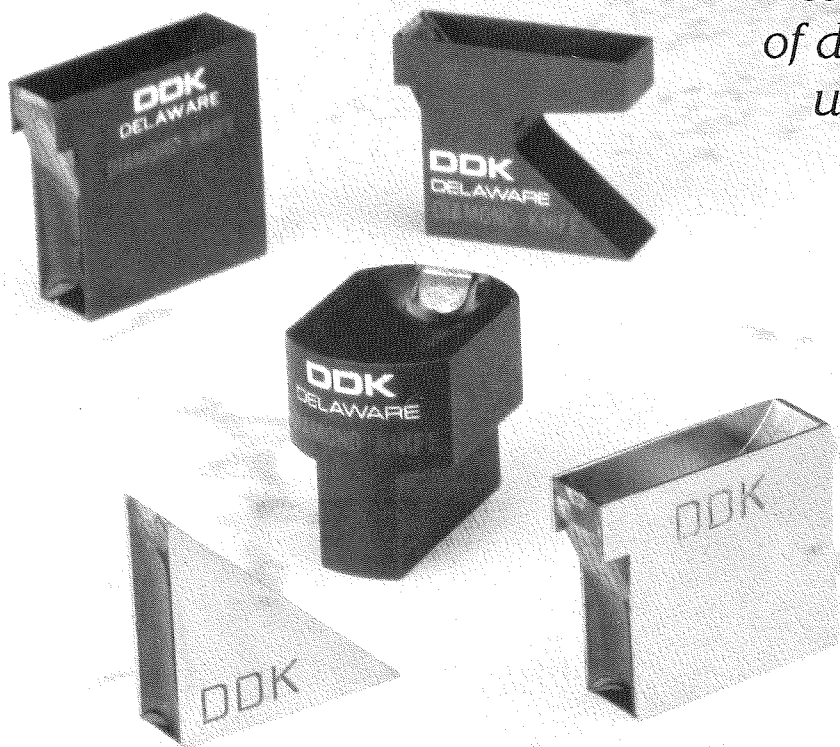
London: MITSUI & CO. UK PLC. (LDNCP Sect.)
20 Old Bailey, London EC4M 7QQ, UNITED KINGDOM

Tel: (71) 822-0597 Fax: (71) 489-0566
Paris: MITSUI & CO. FRANCE S.A. (PRSCC Sect.)
37 Avenue Pierre Premier de Serbie, 75008 Paris,
FRANCE

Tel: (1) 47202761 Fax: (1) 47201908

The Delaware Diamond Knives Family

*Presenting the complete line
of diamond knives for your
ultramicrotomy needs.*



The first step toward perfect images is perfect sections. That's our guarantee to you. Delaware Diamond Knives will give you a great start toward images you'll be proud of.

Whether your work calls for electron or light microscopy, cryo temperatures or ambient, for biological or material samples, DDK has a diamond knife you can depend on. Whether your microtome is fresh from the manufacturer or dusty from the basement, we have a boat style to fit.

We guarantee unlimited resharpenings. Your knife is always restored to its original edge length, cutting angles and like-new condition. You get results, but also that special feeling that comes from owning a quality tool that is also a thing of beauty.

There's no service like DDK service.

Delaware Diamond Knives has long been the innovator in customer service with programs that are the envy of the industry. If we can't treat the customer right, with energy and integrity even when it hurts, we haven't earned your business.

We started the race to shorter delivery times for new knives and sharps. When you absolutely need a good knife now, we offer the only diamond knife replacement program. We still offer custom testing programs and one-on-one ultramicrotomy training in our laboratories. Custom boat styles, colors and special mountings are made in our own shop. Ask about our trade-in program.

DDK's commitment to your satisfaction goes beyond policies. Our unlimited inspection period means that whenever your initial inspection reveals a problem, we will fix it. For the first year, we fix epoxy and mounting problems, regardless of how much you've used the knife. Our warranties are based not on written words but on our commitment to see your work excel.

Call today to see how good customer service can be.



DDK

Delaware Diamond Knives, Inc. □ 3825 Lancaster Pike □ Wilmington, DE 19805
DE: (302) 999-7476 □ US: (800) 222-5143 □ CANADA: (800) 331-3370 □ FAX: (302) 999-8320

Secretary

Texas Society for Electron Microscopy

Center for Biotechnology

Baylor College of Medicine

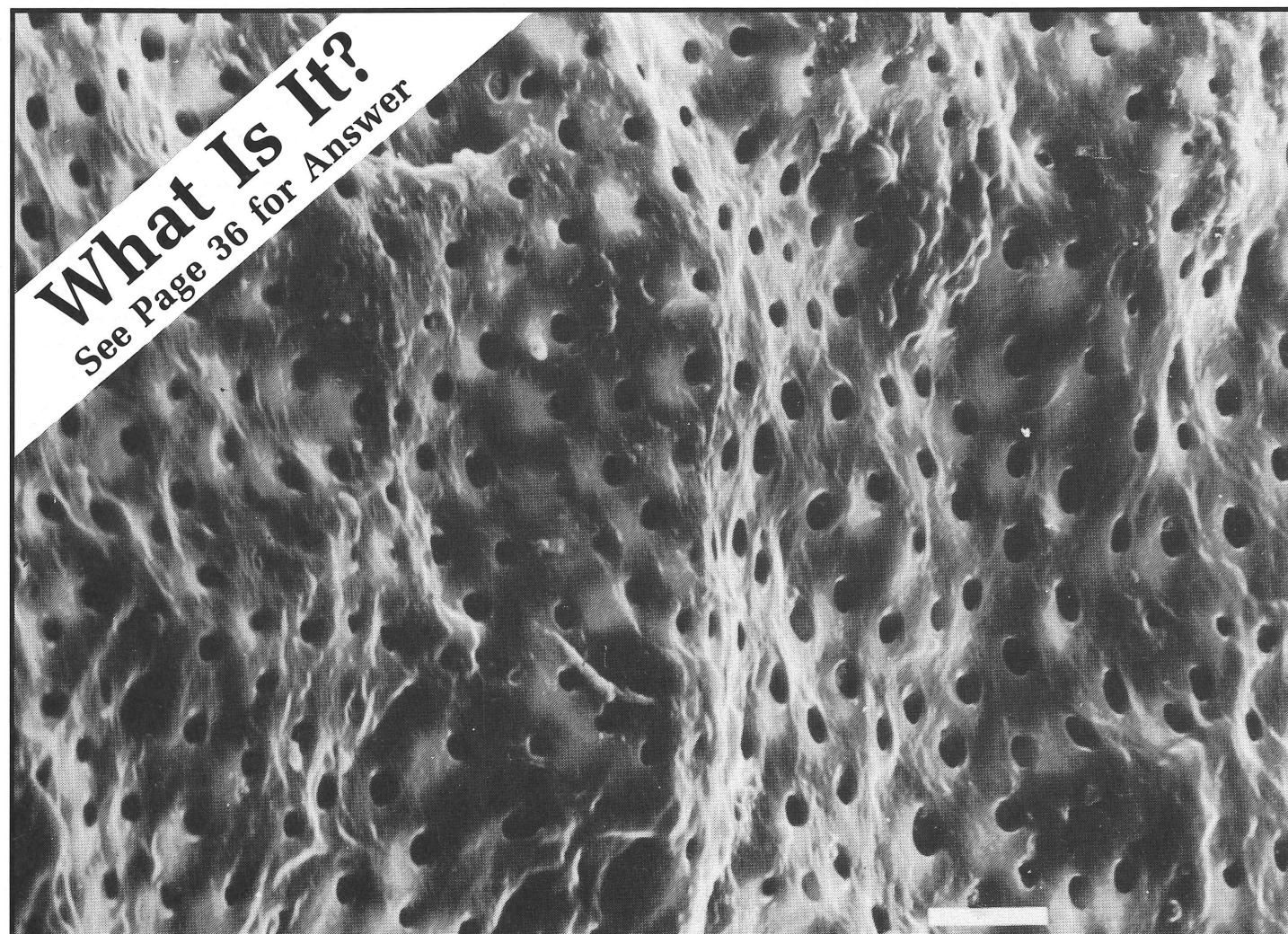
4000 Research Forest Drive

The Woodlands, Texas 77381

(713) 363-8415 • Fax (713) 363-8475

NONPROFIT ORG.
U.S. POSTAGE
PAID
SPRING, TX
PERMIT NO.563

Mannie C. Steglich
Dept. of Pathology - Box 085
Univ. Texas M. D. Anderson Hospital
1515 Holcombe
Houston, TX 77030



Secretary

Texas Society for Electron Microscopy

Center for Biotechnology

Baylor College of Medicine

4000 Research Forest Drive

The Woodlands, Texas 77381

(713) 363-8415 • Fax (713) 363-8416

NONPROFIT ORG.
U.S. POSTAGE
PAID
SPRING, TX
PERMIT NO.563

